# PAKISTAN SUGAR JOURNAL

The first and only research journal regularly published since 1985

#### International Standard Serial Number -1028-1193

#### **ADVISORY BOARD**

Mr. Altaf Muhammad Saleem, Chairman, Shakarganj Sugar Research Institute (SSRI) Jhang, Pakistan	Chairman
Mr. Asad Ullah Khan, Secretary Agriculture, Agriculture Department, Govt. of the Punjab, Lahore, Pakistan	Vice Chairman
Mrs. Rabia Sultan, Member, Punjab Agriculture Commission, Govt. of the Punjab, Lahore, Pakistan	Member
Dr. Ahmad Javed Qazi, Secretary, Industries, Commerce & Investment Department, Government of The Punjab, Lahore, Pakistan	Member
Mr. Nadir Chattha, Secretary, Food Department, Government of Punjab, Lahore, Pakistan	Member
Miss Shakra Jamil, Biotechnologist, Ayub Agricultural Research Institute (AARI), Faisalabad, Pakistan	Member
Mr. Ali Altaf Saleem, Executive Director & Deputy CEO, Shakarganj Limited, Jhang, Pakistan	Member
EDITORIAL TEAM	
Dr. Shahid Afghan, Chief Executive Officer, Sugarcane Research and Development Board, Faisalabad	Editor in Chief
Dr. Naeem A. Gill, Chief Scientist, Sugarcane Research Institute, Faisalabad Pakistan	Member
Dr. Aruna Wejasuria, Sugarcane Research Institute, Dakunu Ala Rd, Udawalawa 70190, Sri Lanka	Member
Mr. Aamir Shahzad, Sugarcane Pathologist, Shakarganj Sugar Research Institute, Shakarganj Limited, Jhang, Pakistan	Editor

Dr. Shahid Afghan, Chief Executive Officer, Sugarcane Research and	Editor in Chie
Development Board, Faisalabad Dr. Naeem A. Gill, Chief Scientist, Sugarcane Research Institute, Faisalabad Pakistan	Member
Dr. Aruna Wejasuria, Sugarcane Research Institute, Dakunu Ala Rd, Udawalawa 70190, Sri Lanka	Member
Mr. Aamir Shahzad, Sugarcane Pathologist, Shakarganj Sugar Research Institute, Shakarganj Limited, Jhang, Pakistan	Editor
Dr. Abdul Khaliq, Department of Agronomy, University of Agriculture, Faisalabad, Pakistan	Member
Dr. Amjad Shahzad, PhD Scholar, PMAS Arid Agriculture University Rawalpindi, Pakistan	Member
Dr. Yong-Bao Pan, Agricultural Research Service (ARS), Department of Agriculture, United States	Member
Dr. William Lee Brusquest, Director, Canavieira Technology Center, Sao Paulo, Brazil	Member
Dr. Jack Charles Comstock, Sugar Cane Growers Cooperative, Belle Glade, Florida, United States	Member
Dr. Phillip Jackson, Commonwealth Scientific and Industrial Research Organization, Canberra, Australia, Australia	Member
Mr. Muhammad Nawaz Khan, DG (R), Ayub Agriculture Research Institute, Faisalabad, Pakistan	Member
Dr. James Todd, Research Geneticist (Plants), Sugarcane Research, United States Department of Agriculture, USA	Member
Mr. Waqas Raza Arshad, Research Officer, Sugarcane Research and Development Board, Faisalabad	Member
Dr. Sagheer Ahmad, National Coordinator Sugar & Food Legume crops, PARC, Islamabad, Pakistan	Member

# **PAKISTAN SUGAR JOURNAL**

#### Open Access Link www.srdb.gop.pk

#### Annual Subscription Rate (4 Quarterly issues)

Pakistan	PKR 1,000/-

Overseas US\$ 100/-

# Published at Shakarganj Sugar Research Institute (SSRI) with the Patronage of Sugarcane Research & Development Board (SRDB)

**Cited by** Asia Net Pakistan (Factiva International Australia) Commonwealth Agriculture & Biology International (CABI-UK)

#### Subscription & Advertisement

M. Ehsan Khan, SRDB, <u>ehsan@srdb.gop.pk</u> Waqas Raza Arshad, SRDB <u>waqas@srdb.gop.pk</u>

#### **International Panel of Referees**

Dr. P. Jackson: Principal Scientist, CSIRO, Australia Dr. Jack C. Comstock: Research Leader, ARS USDA, Canal Point Florida, USA Dr. William Lee Brusquest Director, CTC, Sao Paulo, Republic of Brazil Dr. Raul O. Castillo: Director General, Research Station, Ecuador Dr. Yong-Bao Pan: Research Plant Molecular Geneticist, USDA-ARS, USA Dr. James Todd: Commercial Breeder, USDA-ARS, USA Dr. Niranjan Baisakh: Associate Professor, SPESS, LSU, USA Dr. Arun Wejasuria, Principal Research Officer, Sugarcane Research Institute, Sri Lanka Dr. Peter Allsopp, Editor, International Society of Sugarcane Technologists Dr. Mubashar Nadeem, Assistant Professor, Dept. of Agronomy, UAF Dr. Nazir Javed, Chairman Dept. of Plant Pathology, UAF Dr. Asif Tanvir: Professor, Dept. of Agronomy, UAF Dr. Kashif Riaz, Assistant professor Dept. of Plant Pathology, UAF Dr. Naeem Ahmad Gill, Director, Sugarcane Research Institute, Faisalabad Dr. Abdul Ghaffar, Chairman, Dept. of Agronomy, MNSUAM, Multan Dr. Muhammad Jamil, Post-Doctoral Fellow, KAUST, Saudi Arabia

38

# PAKISTAN SUGAR JOURNAL

CONTENTS	Page
Performance evaluation of Barbados fuzz at initial stages	4
Naeem Ahmad, Muhammad Shahzad Afzal, Dr. Mahmood-Ul-Haasan, Hafiz Basheer Ahmad, Abdul Khaliq, Muhammad Shafique, Naeem Fiaz, Muhammad Akhlaq Muddasar, Muhammad Rizwan Khurshid, Humaira Kausar, Khalid Mehmood Mughal and Naveed Akhtar	
A new approach for integrated pest management of sugarcane in Pakistan	8
Waqas Raza Arshad, Akhtar Hameed and Abdul Khaliq	
Integrated weeds control in sugarcane ratoon management with biotechnological and molecular approaches	16
Naeem Ahmad, Abdul Khaliq, M. Shahzad Afzal, Naveed Akhtar, Waqas Raza Arshad	
<b>Review of Recent Advancements in Sugarcane Genomics</b> Muhammad Ehsan Khan	21
Assessment of Sugarcane Intercropping with different crops Kanza Khan <sup>1</sup> , Muhammad Ehsan Khan <sup>2</sup>	28
SUGAR INDUSTRY ABSTRACTS	33
INTERNATIONAL EVENTS CALENDAR	37

#### **GUIDELINES FOR AUTHORS**

### PERFORMANCE EVALUATION OF BARBADOS **FUZZ AT INITIAL STAGES**

Naeem Ahmad<sup>1</sup>, Muhammad Shahzad Afzal<sup>1</sup>, Dr. Mahmood-Ul-Haasan<sup>1</sup>, Hafiz Basheer Ahmad<sup>1</sup>, Abdul Khaliq<sup>1</sup>, Muhammad Shafique<sup>1</sup>, Naeem Fiaz<sup>1</sup>, Muhammad Akhlag Muddasar<sup>1</sup>, Muhammad Rizwan Khurshid<sup>2</sup>, Humaira Kausar<sup>3</sup>, Khalid Mehmood Mughal<sup>4</sup> and Naveed Akhtar<sup>5</sup> <sup>1</sup>Sugarcane Research Institute, Ayub Agricultural Research Institute (AARI), Faisalabad <sup>2</sup>Oilseeds Research Institute, AARI, Faisalabad <sup>3</sup>Food Technology Post Harvest Research Center, AARI, Faisalabad <sup>4</sup>Statistical Section, AARI, Faisalabad <sup>5</sup>Agronomic Research Institute (V&O) AARI, Faisalabad Corresponding author email: shahzadafzal134@gmail.com

#### ABSTRACT

In this research study, thirteen crosses of sugarcane fuzz received at Sugarcane Research Institute (SRI) Faisalabad from West Indies Central Sugarcane Breeding Station (WICSBS) Barbados were tested and evaluated for initial stages of varietal development program. This study was important as for the first time West Indian sugarcane germplasm was evaluated in Pakistan. The comparison was made with check (approved) varieties to promote into next selection stage. The sowing of fuzz at SRI started during 1<sup>st</sup> week of July 2019 as humidity level and temperature becomes more favorable for its germination. The selection of seedlings was made during the month of November 2019 and 234 clones were promoted to nursery-I. Similarly, 42 clones were advanced to nursery-II. The numbers were reduced tosix in preliminary yield trial when red rot and other disease screening were applied during nursery-II. The clones were planted 1.2m x 1.2m at seedlings stage, 1.2m x 4m at N-I stage, 2.4m x 4m at N-II stage while three replications at preliminary yield trials. The clones showing better performance were carried out to next selection stage of varietal development program during selection process in the month of October each year. Further testing and evaluation of these clones is needed to check more adaptability and stability of general characteristics to proceed into net varietal development stages.

Key words: Sugarcane, Faisalabad. Fuzz, Barbados, Seedling, Selection

#### INTRODUCTION

Sugarcane is vital cash crop that plays vital role in overall GDPA. In Pakistan it falls in top ranked valuable cash crop. The crop has been divide into three zones keeping in view the climatic, soil and other biotic and abiotic factors for successful production of crop. It is cultivated all over the country in irrigated areas. The production of sugarcane stood 81.009 million tones when cultivated on an area of 1.165 million hectare (Govt. of Pakistan, 2021). The climatic

conditions in country do not permit the profuse flowering with enough viability of the sugarcane true seed (fuzz). A specific set of conditions is necessarv to induce the flowering in sugarcane plant ultimately that leads to flowering. A little bit natural conditions exist in Murree that favor flowering in some clones (Ahmed et at., 2019). The changing climatic conditions are disturbing the sugar recovery, growth and overall tonnage of sugarcane crop and causing attack of insects pest, vulnerability to different diseases and lodging

of crop. These challenges urge the investigators to find out well suited clones having good adaptability in different ecological zones. Weather related events and climatic factors play important role to affect yield and production of sugarcane crop (Zhao and Li 2015). In Saccharum officinarum sucrose content range from 12-16% of fresh weight of sugarcane stalk (Bull and Glasziou 1963). Pakistan ranks 5<sup>th</sup> position by area and production among producina world sugar countries (FAO, 2017).

Genetic variability is important

for developing new variety of any crop. Fuzz provide the opportunity to develop new variety in sugarcane as crossing over creates genetic variability at meiosis level during seed formation. The researchers try to fine better improved sugarcane and clones that have high sugar contents and good tonnage that are more profitable for growers and millers (Jackson, 2005). Bringing desirable parents in cross combinations during hybridization process is key to evolve new crop The variety. vield being quantitative is closely linked with environmental factors (Pandey et al. 2018).

Sugarcane Research Institute, Faisalabad Pakistan working has been on sugarcane varietal development program for many decades. Normally the sources of fuzz have been USA, Australia, South Africa, Brazil and Barbados for evolution of sugarcane varieties. However, a few varieties from local fuzz were also evolved but main source of varieties development have been exotic fuzz or introduction. Sugarcane Research and Development Board, Pakistan being а funding agency, have been providing fuzz to Sugarcane Research Institute. Faisalabad from last many Durina 2018-19. vears. thirteen crosses of Barbados were provided to be included in varietal development program. The purpose of current study is to find out best suited clones having more adaptability in local climatic conditions and having better or comparable performance with existing check varieties.

#### **Plantation of Experiment**

The experiment was planted at coordinate plane 31.4311° N and 73.0694° E during 2018, 2019 and 2020 at research farm of Sugarcane Research Institute, Faisalabad. The soil texture was loamy having electric conductivity (EC) 1.85, soil pH 7.9, organic matter 0.95%, available phosphorous 8.4 ppm, available potassium 189 ppm and saturation 30 %.

#### Seedlings Stage

The fuzz was sown during July, 2018 on raised beds. For proper germination of sugarcane fuzz requires relative humidity more than 60 % and temperature range from 30 to 38C. As monsoon season starts. the required temperature and humidity goes up to mark and provides favorable condition for good germination of fuzz. Thirteen crosses of Barbados were of sown and all them aerminated providing 4077 The seedlings. seedlings were singled out in earthen pots and transplanted into field at the arrival of favorable environment when frost ended. During October 2019, selection of seedlings was made taking brix value in comparison to check varieties. During selection process 234 clones were selected and promoted to N-I stage for further evaluation at single line stage. The brix % of HSF 240 and CPF 249 stood 18 and 19 respectively. While the brix and other characters of each stool were compared with check varieties and promoted to next stage for further evaluation (Table-1).

#### Nursery -1

selected The clones in seedlings stage were planted in single row of Nursery one stage along with three check varieties i.e HSF 240, CPF 249 and CPF 253 during October 2019. In this stage one row of 4 meter is planted by each promoted clone. Among 234 clones, 42 clones were found good and promoted to next stage called "Nursery-II" (Table-2). The selection index at this stage 18%. All other remained clones were not found good and eliminated from varietal development program. Liberal selection was made at this stage. The selection process of this stage was carried out during October, 2020. This stage is actually seed increase stage here only single replication is used.

#### Nursery-II

Nurserv-II is double row single replication stage. The plot size of each entry kept 2.4m x 4m. At this stage 42 clones along with three check varieties were tested. Selection was made during October, 2021 to check the performance of these clones comparison to in check varieties. At this stage the reaction to red rot pathogen was also checked. The clones that showed susceptibility to red rot, showed lodging, high pithy, smut disease, poor growth and stand were rejected and only 6 clones were proved good according to selection criteria and

#### PSJ APRIL-JUNE, 2022 ISSUE

Vol. XXXVII, No.02

promoted Nursery-III to (Preliminary Yield Trial). These clones have been planted to Nursery-III stage using three replications deploying Randomized Complete Block Design (RCBD). Recommended seed rate along with recommended cultural practices were applied to evaluate at this stage (Table-3). The plot size for single replication is kept 3.6m x 4 m. while promoting clones from Nursery-I to Nursery-II the selection index remained 14%. These clones are under study current year and their performance will be evaluated at harvest time, January 2023. The clones showing good performance will be carried to Semifinal stage.

Sr. No. Code Parentage		No. of Seedlings	Selected Clones	Brix (%)	Clone No. Allotment	
1	1 9 D 98 398 x Poly CC		312	6	17-18	BDF 19-(16)
2	10	CR 10 2007 x Poly CC	342	35	16-22.5	BDF 19-(741)
3	2	BR 10 001 x Poly CC	216	4	15-22	BDF 19-(4245)
4	4	CR 08 0014 x Poly CC	343	29	16-21	BDF 19-(4674)
5	11	CR 10 2003 x Poly CC	229	20	17-22	BDF 19-(7594)
6	5	B 09 2681 x Poly CC	290	17	16-22	BDF 19-(107123)
7	1	CP 96 1252 x CR 11 0004	130	7	15-20	BDF 19-(124130)
8	6	BJ 06 794 x Poly CC	230	27	17-22	BDF 19-(131157)
9	7	BR 08 008 x Poly CC	420	12	16-22	BDF 19-(158169)
10	8	D 93 224 x Poly CC	298	10	17-19	BDF 19-(170179)
11	3	DB 04 01 x Poly CC	312	1	19	BDF 19-(180)
12	12	BJ 99 112 x Poly CC	940	66	15-25	BDF 19-(181246)
13	13	DB 04 1009 x Poly CC	15	0	-	-
	Total		4077	234		

#### Table-1 Selected Clones in Seedlings to Promote in Nursery-1

#### Table-2 Clones Promoted to Nursery-II from Nursery-I

Sr. No.	Clone No.	Brix (%)	Sr. No.	Clone No.	Brix (%)	Sr. No.	Clone No.	Brix (%)
1	BDF 19-1	15.3	16	BDF 19-41	16.3	31	BDF 19-112	18.3
2	BDF 19-7	16.7	17	BDF 19-45	20.7	32	BDF 19-139	21.3
3	BDF 19-10	18.7	18	BDF 19-48	20.3	33	BDF 19-146	19.3
4	BDF 19-11	18.7	19	BDF 19-50	19.0	34	BDF 19-177	18.3
5	BDF 19-12	20.7	20	BDF 19-52	17.0	35	BDF 19-181	21.0
6	BDF 19-15	19.3	21	BDF 19-54	18.0	36	BDF 19-186	19.0
7	BDF 19-16	20.0	22	BDF 19-56	17.3	37	BDF 19-205	20.7
8	BDF 19-22	18.3	23	BDF 19-67	20.3	38	BDF 19-223	17.7
9	BDF 19-23	19.3	24	BDF 19-70	19.0	39	BDF 19-227	15.0
10	BDF 19-24	18.3	25	BDF 19-77	18.0	40	BDF 19-230	19.7
11	BDF 19-25	17.3	26	BDF 19-79	15.7	41	BDF 19-245	19.3
12	BDF 19-28	19.0	27	BDF 19-92	19.3	42	BDF 19-246	18.3
13	BDF 19-29	19.3	28	BDF 19-102	19.3	43	HSF 240	19.3
14	BDF 19-31	17.3	29	BDF 19-105	19.7	44	CPF 249	19.0
15	BDF 19-35	18.0	30	BDF 19-110	21.0	45	CPF 253	19.7

#### Table-3 Clones promoted to Nursery-III (Preliminary Yield Trials)

Sr. No.	Clones	Brix (%)	Sr. No.	Clones	Brix (%)
1	BDF 19-25	17.7	6	BDF 19-230	16.0
2	BDF 19-45	15.7	7	HSF 240 (C1)	19.3
3	BDF 19-54	16.0	8	CPF 249 (C2)	18.0
4	BDF 19-92	16.3	9	CPF 253 (C3)	17.3
5	BDF 19-102	17.0			

#### Vol. XXXVII, No.02

#### CONCLUSION AND RECOMMENDATIONS

The varietal development program adapted at Sugarcane Research Institute, Faisalabad includes both fuzz and introduction for testing, evaluation, selection and promoting the clones to develop genetically improved sugarcane varieties. The 1<sup>st</sup> way is extensively used for developing sugarcane varieties. Sowing of fuzz in July gives good germination results while its selection during the month of October is favorable when temperature lowers and brix percent starts to increase. Six clones BDF 19-25, BDF 19-

45, BDF 19-54, BDF 19-92. BDF 19-102 and BDF 19-230 from Barbados fuzz are under evaluation and will be promoted to semifinal for further study. The best performing clones may be released as variety while checking all required aspects in latter stage of varietal development program.

#### REFERENCES

Ahmed M. F., M. Siddique, N. Kamal and D. N. Ahmad. 2019. Sugarcane flowering at sugarcane breeding sub-station (SBSS), Murree. Haya S. J. of Life Sci., 206-212.

Bull TA and K.T. Glasziou. 1963. The evolutionary significance of sugar accumulation in *Saccharum*. Aust. J. Biol. Sci., 16: 737–742.

FAO, 2017. Food and Agriculture Organization of the United Nations. Available at: http://www.fao. org/faostat/en/#data/QC (Accessed: 20 October 2019).

Govt. of Pakistan, 2021. Pakistan Economic Survey: Agriculture. pp: 22.

Jackson, P. A. 2005. Breeding for improved sugar content in sugarcane. F. Crop. Res. 92(2–3): 277–290.

Pandey D, SP Singh, A. S. Jeena, K. A. Khan, T.A. Negi D. Koujalagi. 2018. Study of genetic variability, heritability and genetic advance for various yield and quality traits in sugarcane genotypes (*Saccharum offiinarum*). Int. J. Curr. Microbiol. App. Sci., 7:1464–1472.

Zhao, D., and Y.-R. Li. 2015. Climate change and sugarcane production: potential impact and mitigation strategies. Int. J. Agron. 2015.

### A NEW APPROACH FOR INTEGRATED PEST MANAGEMENT OF SUGARCANE IN PAKISTAN

Waqas Raza Arshad\*, Akhtar Hameed\*\* and Abdul Khaliq\*\*\* \*Sugarcane Research and Development Board Punjab Pakistan \*\*Muhammad Nawaz Sharif University of Agriculture Multan Pakistan \*\*\*Sugarcane Research Institute Faisalabad Pakistan

#### ABSTRACT

Conventional Integrated Pest Management (IPM) is mostly described as "a decision-making process using multiple pest management tactics to prevent economically damaging out-breaks while reducing risks to human health and the environment". Low-level IPM is the most often employed form, consisting of the most basic of IPM practices-scouting and insecticide applications according to economic thresholds. Some growers have progressed to medium-level IPM, the adoption of a few additional preventive measures, e.g. cultural controls and plant resistance, coupled with efforts to cut back on broad spectrum pesticide use in order to protect beneficial organisms. These IPM strategies are mainly targeted towards single pest species and do not consider all the pests in a specific agro-ecosystem. High-level or Bio-intensive IPM, is where multiple interventions are integrated in a bio-intensive approach targeting multiple pests. Bio-intensive IPM is based on holistic agro-ecosystem interactions, in which knowledge about insects, their symbionts, pathogens, natural enemies, plants, endophytes and interactions between all of these are combined to develop IPM in an area-wide, environmentally friendly manner. Reviewed here are advances in knowledge of, and of biotic interactions between direct, indirectand induced plant resistance, plant nutrition, habitat management, chemical ecology, natural enemies, soil-health, micro- organisms such as endophytic fungi and Wolbachia and phylogenetics and phylo-geography. All of these are potential building blocks of a bio-intensive IPM system under-construction at SRDB, SRI and MNSUAM. Also discussed are opportunities and challenges in these areas of research, considering bio-security threats to the Pakistan sugar industry and possible limitations in current sugarcane plant breeding material.

**Keywords:** *Chiloinfuscatellus*, IPM, Induced Resistance, Sugarcane, Pakistan, Direct and Indirect Resistance.

#### INTRODUCTION

SRI Faisalabad has been working to improve control of the sugarcane stem borer Chiloinfuscatellus (Snellen) since the early 2010s (Munir, 2014). A few cultural control measures and several less susceptible varieties have been developed against it (Munir, 2014). However, it still remains a pest throughout the sugar industry (Sikandar and Ahmad, 2021).

In order to build resilience

into the sugarcane agroecosystem, a refocusing of control efforts into a biointensive area-wide integrated pest management approach necessarv is (Klassen, 2005). Such an approach marries conventioncontrol options with al ecologically based new technologies such as delineation of within species populations, chemical ecology, stimulo-deterrent diversion (push-pull) and enhancement natural of enemies through habitat

management and good soil health practices, to produce sustainable IPM strategies applicable across large areas involving multiple stake (Conlong holders and Rutherford, 2009).There is also a need to refocus biosecurity build to again resilience to invasion into agro- ecosystems, rather than building walls around them.

#### **IPM–From the bottom up**

Plant resistance to pests and diseases can be linked to optimal physical, chemical

#### PSJ APRIL-JUNE, 2022 ISSUE

and biological properties of soil (Zehnder et al., 2007). 'Healthy' soil is described as sufficient having organic matter to support a high diversity of animal (arthropods, nematodesetc.) and microbial life. Soil can act as important reservoirs for a diversity of entomo pathogenic fungi & nematodes. well as as predaceous arthropods. which contribute can significantly to the regulation of pest populations.

Ninety percent of insect pest species spend at least part of their life cycle in soil. In addition. pests that occasionally come into contact with soil can be attacked by predators or become infected by entomo pathogens (Klingen et al., 2002). After the harvest of heavily infested sugarcane, the residual E. saccharina population from which infestation of the following ratoon crop can be expected is found in the sugarcane stubble at soil level and in the stool below ground.

Bv minimizing compaction and tillage, and by mulching organic increasing and matter, soil scan support increased populations of pathogenic fungi, entomo pathogenic entomo nematodes and predaceous arthropods such that these natural enemies of insects can be included in the suppression of pests in a conservation biological control strategy (Meyling and Eilenberg, 2007).

Direct and indirect host-

#### plant resistance

Insect resistance in grasses is the result of many defense mechanisms that act in parallel to limit the damage of herbivore attacks. Many of these defense mechanisms are based on plant secondary metabolites, or defensive proteins that directly affect the herbivore due to their toxic, deterring or anti-nutritional properties.

Structural resistance also occurs. Keeping and Meyer (2002) have shown that resistance to E. saccharina can be enhanced using soilapplied silicon. which becomes incorporate in to the plant alongside lignin and fiber increasing resistance to penetration. These authors emphasize relationship а between nitrogen and silicon nutrition where by the ratio of these elements determined in leaf analyses can be used as an indicator of E. saccharina infestation risk. Keeping and Rutherford (2004)have reviewed mechanisms of Ε. direct resistance to Two decades saccharina. ago, a new type of defense mechanism, termed indirect defense, was first described in maize. Central to this type of defense is the release of a volatile plant SOS signal, a mixture of volatile secondary metabolites.

Plant volatiles are derived from complex biochemical processes and include fattyacid-derived products [methyl-jasmonate, cisjasmone, and green leaf volatiles (GLV) like hexenal and hexenyl-acetate], monoterpenes. sesquiter

and shikimicacidpenes. derived products [e.g.methylsalicylate and indole] (Ferry et al., 2004). These can serve as signals, not only to attract predators and parasites of attacking herbivores, but they can also repel the herbivore itself, and they can elicit responses in neighbouring undamaged plants (De Moraes et al., 2001). The use of elicitors to directly activate or prime resistance shows much promise as an IPM tool (Zehnder et al., 2007).

#### Habitat management

Therefore, it is very important to understand the role of plants in managing in sect populations. An example comes from our experience in trying to control E. saccharina with indigenous association and new biological control agents.

Conlong *et al.* (2007) found that female *E. saccharina* moths will accept *Cyperus papyrus* and *Cyperus dives* as host plants in preference to the indigenous grass *Pennisetum purpureum*, with sugarcane being least preferred.

A preference was demonstrated by Keeping *et al.* (2007), that if given choice between older sugarcane & maize, *E. saccharina* would ovi position maize even if it were Bt maize.

Keeping *et al.* (2007) further showed that larval survival on this Bt maize was zero. A hierarchicalovi position preference (Thompson and Pellmyr, 1991) is suggested in Southern African *E*. saccharina females, with most ovi position found on or close to its indigenous sedge hosts, followed by indigenous grasses, and then sugarcane.

However, a large proportion of these eggs were not laid directly on the plants, but in crypticovi position sites in the vicinity of potential host plants (Kasl, 2004; Barker, 2008). Egg dumping is behavior of highly poly phagous species (E. saccharina attacks species of the Cyperaceae, Typhaceae, Juncaceae and Gram in (Conlong, 2001: aceae Conlong, Mazodze and 2003), orin species associated with super abundant host plants.

These are both possibilities with Ε. saccharina in its sugarcane and cyperaceous hosts, as both hosts occur large in essentially mono-specific stands. Adult females therefore not be may particularly attracted by host or 'pull' plants in an IPM system and conversely they may be more strongly repelled by non-host or 'push' plants, since the presence of these could indicate that the insect had reached the edge preferred of the monospecific host plant stand.

Nevertheless, *E. saccharina* seems to have a hierarchical preference in choosing a host plant habitat to oviposit in, i.e. Cyperaceae and maize, both of which have *E. saccharina* population controls in place; natural enemies in the Cyperaceae (Conlong, 1990, 1997, 2000) and genetically engineered Bttoxininmaize (Keeping *et al.*, 2007).

Further evidence to promote habitat management as a control option, demonstrated the repellent 'push' properties the indigenous of grass Melinisminutiflora Beauv. to cereal stem borers, and also its attractant properties to parasitoids. their М. minutiflora produces volatiles similar to damaged maize, even in the absence of pest damage to itself (Gohole et al., 2003).

In a glasshouse experiment at SASRI. Xanthopim plastemmator (Thunberg) (Hymenoptera: Ichneumonidae) parasitised more E. saccharina pupae in sugarcane in close proximity grass, than to this in sugarcane only (Figure 1) (Kasl, 2004). This suggests that the searching behaviour of the parasitoid was increased by Melinis volatiles.

The next phase in developing this habitat management approach for E. saccharina was to setup field trials using rows of M. minutiflora along either in irrigation or contour breaks as a repellentor 'push' saccharina plant. Ε. populations and damage were halved in field plots planted next to strips of M. minutiflora compared to control plots, suggesting that the pest was repelled by Melinis volatiles (Figure 2) (Barker et al., 2006). Planting Cyperus papyrus as a trap, or 'pull' plant along drainage

lines of selected sugarcane fields resulted in significantly reduced damage in the cane associated with it (Figure 3) (Kasl, 2004).

Based on the success of these trials, a farm-based habitat management plan has been devised. incorporating indigenous host plants and Bt maize as 'pull' plants for E. saccharina and M. minutiflora as the 'push' component. This biointensive approach has been expanded into Bioа intensive-PM plan, in corporating plant nutrition, soil health and the use of less susceptible sugarcane varieties.

An addeda spectto the plan is to plant buck wheat at the time of sugarcane planting. This is to attract adult parasitoids and predators into the sugarcane environment by providing a pollen and nectar source for their survival during periods of low host availability, much the same as advocated by Wäckers et al. (2005) and Zehnder et al. (2007) in their conservation biological control approach to enhance the activity of indigenous natural enemies.

# Does sugarcane emit SOS volatiles when attacked by *E. saccharina*?

In contrast to the situation in the natural hosts of *E.* saccharina, negligible parasitism has been recorded in sugarcane, even when this crop was planted adjacent to infested indigenous host plants with a bundant parasitoids present (Conlong and Hastings, 1984). Many introduced parasitoid shave also failed to colonise the sugarcane habitat (Conlong, 1997).

Using gas chromatography, Smith et al. (2006) showed different volatile emission patterns between Cyperus infested bv E. papyrus saccharina and un-infested С. Infested papyrus. sugarcane was neither qualitatively or quantitatively different from un-infested sugarcane and both were different from C. papyrus (Figure 6). In addition, these authors showed that the parasitoid Goniozusindicus (Hymenoptera: (Ashmead) Bethylidae) was attracted to frass from E. saccharina that had fed on C. papyrus, and was not attracted to frass from E. saccharina that had fed on sugarcane. Adding this to the lack of parasitism recorded in sugarcane, even in the vicinity of natural host plants harbouring parasitoids, modern suggests that sugarcane genotypes may not attract natural enemies through the release of herbivore induced SOS volatiles, or that they may differ in the ability to do so.

## Genotypic differences in plant volatile emission

The ability to mount indirect defence against Ε. saccharina may have been lost in sugarcane as a result of in advertently concentrating direct on resistance in amono culture oriented breeding plant selection program. Besides this possibility, the release of

volatiles plant is characterised bya large degree of genotypic variation plant species, within for example, maize genotypes and their closest wild relatives, Zea mays ssp. Parviglumis and Mexicana (collectively known as teosinte), show significant differences in emissions when attacked (Gouinguene et al., 2001; Degen et al., 2004).

An example of loss of indirect defence has been found below ground in maize. In response to feeding by the western corn rootworm. virgiferavirgifera Diabrotica (LeConte) (Coleoptera: Chrysomelidae), maizeroots release (E)-b-caryophyllene that attracts the entomo pathogenic nematode Heteror habditismegidis (Rasmann et al., 2005). Most North American maize lines do not release (E)-bcaryophyllene in response to rootworm attack, whereas many European lines and teosinte accessions do (Kollner et al., 2008).

The existence of genotypic differences in the emission pattern of volatile compounds for Kenvan M. minutiflora cultivars has also been demonstrated (Goholeet al., 2003). The lack of response by X. stemmator in the presence of Australian M. minutiflora again points to variability within this species (Figure 1). Australian M. minutiflorais extensively used for cattle fodder.

The strong od our of the plant can be carried through to milk and, because of this, there has been an extensive program to breed less volatile variety with similar nutritional quality. The volatile (s) that the parasitoid responded to in the African variety could have been bred out of the Australian variety.

#### Breeding for artificially primed and induced resistance

The loss of the ability to produce an SOS volatile and the observed genotypic variability in their production by maize points towards the exploitation of the phenomenon in sugarcane, by breeding varieties for enhanced attractiveness to natural enemies. This could achieved through the be application of an artificial elicitor followed by selection enhanced direct and for indirect resistance in а system that includes natural enemies.

Experimental application of elicitors is fairly simple and it is worth trying to make selections among plant breeding lines grown under the influence of plant defence elicitors. aiming for new cultivars optimized for artificially inducible resistance without traits significant yield penalty (Agrawal et al., 2002; Ahman, 2006). Historically, induced resistance research has mostly concentrated on direct activation where resistance is expressed in advance of challenge by the pest. The possibility of priming as a mechanism of protection has been overlooked often because it only becomes in challenged apparent

Vol. XXXVII, No.02

plants. Priming equates to a 'heightened state of readiness', in that in the event of damage to a primed plant, resistance responses are faster and more intense (Conrath *et al.*, 2006).

Direct activation of resistance might best be employed where the target pest is widespread and has predictable outbreaks. An example is the sugarcane Fulmekiolaserrata thrips, (Kobus) (Thysanoptera: Thripidae) that affects more than two thirds of sugarcane plants in a particular field at the same time. Outbreaks summer occur in with numbers peaking every January since the pest was first discovered on the African continent in 2004 (Way et al., Primed resistance 2006). would, however, be more suitable for E. saccharina, much lower since а plants proportion of is infestation attacked and stend to be patchy.

# Ecology and phylo geography

The basic building block of IPM is stiller garded as ecology (Gurr et al., 2003). In a study of E. saccharina, Conlong (2001) found behavioural, host plant and natural enemy differences in population soccurring between South. Central and Africa. with West them seemingly coming together in Uganda.

The second fusing factors between different populations of what is otherwise amorphologically similar

species made it an ideal can did at e for molecular systematic canalyses. Assefa et al. (2006), usina the cytochrome oxidase subunit 1 (CO1) region of the mitochondrial genome, separated E. saccharina into three distinct groups (west, south and Ethiopian). Two of these groups (west and south) were found in Uganda. The CO1 genetic diversity between these groups was larger than that between recognised species within the genus Ostrinia (Lepidoptera: (Coatesetal., Crambidae) 2005). In other insects, unexpected mt DNA CO1 patterns have led to the discovery of cryptic species (Hebert et al., 2004; et al., 2007). Such diversity should encourage us to confirm covarying genetic, behavioural and ecological characteristics which would lend support to the notion that cryptic species exist within the E. saccharina complex.

In IPM programs which use classical bio-control as one of their management options, or translocation of natural enemies, these aspects can be enhanced by using such techniques to identify cryptic species, or populations of species most closely related to each other, so that more informed decisions can be made regarding natural selection for enemy use against pests. This applies not only to pest species, but also to parasitoids (Ngi-Song et al., 1998).

Since 1992, surveys for indigenous parasitoids of *E*.

saccharina in a variety of African habitats have been completed. Thirty species of larval parasitoids have been found in eight countries (Conlong, 2000). Several of have failed these to parasitise E. saccharina from South Africa due to incompatibility. For example, West from Africa. Descampsinasesamiae (Diptera: Tachinidae) larva eareen capsulated by E. saccharina (Conlong, 1997). Cotesiasesamiae (Cameron) (Hymenoptera: Braconidae) from South A fricaisalsoun successful as sareen capsulated. egg Further collections of parasitoids from Central Africa, where southern and western populations of E. saccharinaco-exist, could revealbiotypes of parasitoids that could be effective against this pest (Ngi-Song et al., 1998).

## The interaction between *E.* saccharina and *Fusarium*

When E. saccharina bores in sugarcane, the tissue surrounding the borina becomes reddish discoloured often affecting а whole internode. Following on from this, and work of Schulthess et al. (2002), McFarlane et al. (2009) cultured numerous Fusarium spp. isolates, from the red tissue surrounding E. saccharina borings, as well as from undamaged stalks as endophytes. Most of the isolates from borings were beneficial to E. saccharina in artificial diet in terms of larval survival and growth rate, and attractive were to neonatesinol factory choice assays.

A few of the endophytic isolates were antagonistic, with *E. saccharina* neonates repelled and growth retarded. Moths may also be attracted or repelled depending upon isolate. Ako *et al.* (2003) showed that West African *E. saccharina* females laid on average 32 eggs on maize stalks with *F. verticillioides* present as an endophyte, versus nine on stalks grown from fungicide or hot water treated seeds.

integrated control In an approach against Ε. saccharina, seed cane hot water treatment and/ or treatment with fungicides endophytic could reduce colonisation bv Fusarium beneficial to E. isolates saccharina, thereby reducing the chance of infestation. Alternatively, the facilitation of endophytic colonization of sugarcane by Fusarium isolates antagonistic to E. saccharina could afford more sustainable and environmfriendly protection entally from this stalk borer. Another approach could be to exploit the differences in volatiles between repellent and isolates the attractive in development of repellents and lures of use in the field.

#### CONCLUDING REMARKS

Khan et al. (1997a) described a 'push-pull' system effective against stem borers in sugarcane. This system indigenous includes the Cotesiasesamiae parasitoid as well as C. flavipes which were introduced against the Chilopartellus exotic

(Overholt et al., 1997). C. sacchari phagus, now present in Mozambique, is being parasitised by both C. flavipesand X. stemmatoron larvae and pupae respectively in sugarcane. Ngumbi et al. (2005) showed that C. flavipes females respond to terpenoids and the green leaf volatiles which are released by maize plants damaged by C. partellus. This suggests that sugarcane may produce SOS volatiles in response to С. sacchariphagus.

There are alternative hypotheses to that of loss of ability to produce SOS volatiles. By boring the tops of stalks and leaf spindles in young cane, volatile emission could be elicited by C. sacchariphagus, whereas E. saccharina may avoid eliciting volatile emission by boring the bottoms of stalks in older cane. Another hypothesis that is the Fusarium associated with E. saccharina could interfere with the elicitation of volatile emission or change the of emitted composition volatiles. These possibilities worthy are of further investigation.

Nevertheless, a bio-intensive IPM strategy could be made more effective if the crop itself is capable of releasing appropriate SOS volatiles attacked. Breeding when for enhanced varieties attractiveness natural to enemies potential if has natural enemies are already habitat present in the management system. Goniozusindicus parasitises

E. saccharina in C papyrus. It is also known to parasitise C. partellus (Keiji and Overholt, 1996) and has been found doingso in Sorghum growing in arundinaceum C. papyrus proximity to (Conlong, 1994; 1997). It is therefore possible that G. indicus could parasitise both C. sacchariphagus and E. saccharina in sugarcane. The same applies to the pupal parasiteX. stemmator.

Conlong et al. (2004) showed that some of the varieties with the highest direct resistance to E. saccharina were the susceptible С. most to sacchariphagus and viceversa. This has implications for variety choice in an IPM system designed to target both pests simultaneously, sacchariphagus should С. invade the South African industry. Given sugar increasing adoption of the 'push-pull' habitat management concept even in the absence of parasitoid activity against E. saccharina in sugarcane, and its success against C. partellusin maize, we are confident that 'pushpull' habitat management will also be effective against C. sacchariphagus.

The development of IPM strategies depends on a sound understanding of the chemical ecology of pest interactions with sugarcane. natural enemies and the habitat. Modern IPM is not only about insect/plant interactions, it about is holisticagro-ecosystem interactions. in which increased knowledge about the environment. soils.

plants, pathogens, endophytes, symbionts and insects are all combined to provide effective crop protection in an environmentally friendly manner. As knowledge about, and interactions between, induced resistance. plant chemical ecology, microorganisms such as endophytic fungi and Wolbachia, and phylo genetics and phylo geography of arthropods becomes easily available; it is hypothesized that these will become important components of bio-intensive AW-IPM, thereby minimizing the impacts of synthetic pesticides even more.

#### REFERENCES

Ahman, I. (2006). Breeding for inducible resistance against insects–applied plant breeding aspects. In: Abstracts of the IOBC Meeting, Breeding for inducible resistance against pests and diseases.

Assefa, Y., Mitchell, A. and Conlong, D.E. (2006). Phylo-geography of Eldanasaccharina Walker (Lepidoptera: Pyralidae). Ann. Soc. Entomol. Fr., 42: 331–337.

Barker, A. J. (2008). Habitat management using stimulo deterrent diversion techniques to decrease infestation of sugarcane by Eldanasaccharina Walker (Lepidoptera:Pyralidae) Unpublished MSc Thesis. Faculty of Science, University of the Witwatersr and, Johannesburg, South Africa.128 p.

Barker, A. J., Conlong, D. E. and Byrne, M. (2006). Habitat management using Melinisminutiflora (Poaceae) to decrease the infestation of sugarcane by Eldanasaccharina (Lepidoptera: Pyralidae).Proc. S. Afr. Sug. Technol. Ass., 80: 226–235.

Coates, B. S., Sumerford, D. V., Hellmich, R. L. and Lewis, L. C. (2005). Partial mitochondrial genome sequences of Ostrinianubilalis and Ostriniafurnicalis. Int. J. Biol. Sci., 1:13–18.

Conlong, D. E. (2000). Indigenous African parasitoids of Eldanasaccharina (Lepidoptera: Pyralidae).Proc. S. Afr. Sug. Technol. Ass., 74:201–211.

Conlong, D. E. (2001).BiologicalcontrolofindigenousAfricanstemborers:Whatdoweknow? Insect Sci. Applic., 21: 267–274.

Conlong, D. E., Sweet, P. and Piwalo, J. (2004). Resistance of southern African varieties of sugarcane to Chilosacchariphagus (Lepidoptera: Crambidae) in Mozambique, and development of a non-destructive field resistance rating system. Proc. S. Afr. Sug. Technol. Ass., 78: 297–306.

Conlong, D. E., Kasl, B. and Byrne, M. (2007). Independent kids-or motherly moms? Implications for integrated pest management of Eldanasaccharina Walker (Lepidoptera: Pyralidae). Proc. Int. Soc. Sugar Cane Technol., 26: 787–796.

Cook, S. M., Khan, Z.R. and Pickett, J. A. (2007). The use of 'push–pull' strategies in integrated pest management. Ann. Rev. Entomol., 52: 375–400.

De Moraes, C. M., Mescher, M.C. and Tumlinson, J. H. (2001). Caterpillar-induced nocturnal plant volatiles repel con specific females. Nature,410: 577–580.

Ferry, N., Edwards, M. G., Gatehouse, J. A. and Gatehouse, A. M. R. (2004). Plant-insect interactions: Molecular approaches to insect resistance. Curr. Opin. Biotechnol., 15: 155–161.

Gohole, L. S., Overholt, W. A., Khan, Z. R., Pickett, J. A. and Vet, L. E. M. (2003). Effects of molasses grass, Melinisminutiflora, volatiles on the foraging behavior of the cereal stem borer parasitoid, Cotesiasesamiae. J. Chem. Ecol., 29: 731–745.

Gurr, G. M., Wratten, S. D. and Altieri, M. A. (eds) (2004). Ecological Engineering for Pest Management. Advances in Habitat Manipulation for Arthropods. CSIRO Publishing, Collingwood, Victoria 3066, Australia.

Hebert, P.D.N., Penton, E.H., Burns, J M., Janzen, D. H. and Hallwachs, W. (2004). Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly Astraptesfulgerator. Proc. Natl Acad. Sci. USA, 101: 14812–14817.

Keeping, M. G. (2006). Screening of South African sugarcane cultivars for resistance to the stalk borer Eldan asaccharina Walker (Lepidoptera: Pyralidae). Afr. Entomol., 14: 277–288.

Keeping, M. G. and Rutherford, R. S. (2004). Resistance mechanisms of South African sugarcane to the stalk borer Eldan asaccharina (Lepidoptera: Pyralidae): A review. Proc. S. Afr. Sug. Technol. Ass., 78: 307–311.

Keeping, M. G., Rutherford, R. S. and Conlong, D. E. (2007). Bt maize as a potential trap crop for management of *E.saccharina* Walker in sugarcane. J. Appl. Entomol., 131: 241–250.

Klassen, W. (2005). Area-Wide Integrated Pest Management and the Sterile Insect Techniques. In Dyck, V. A., Hendrichs, J. and Robinson, A. S. (Eds). Sterile Insect Technique. Principles and Practice in Area-Wide Integrated Pest Management. pp 39–68. IAEA Springer. The Netherlands.

Klingen, I., Eilenberg, J. and Meadow, R. (2002). Effects of farming system, field margins and bait insect on occurrence of insect pathogenic fungi in soils. Agric. Ecosystems Environ., 91: 191–198.

Kollner, T. G., Held, M., Lenk, C., Hiltpold, I., Turlings, T. C. J., Gershenzon, J. and Degenhardt, J. (2008). Amaize (E)-b-caryophyllenesynthase implicated indirect defense responses against herbivores is not expressed in most American maize varieties. Plant Cell, 20: 482–494.

Mazodze, R. and Conlong, D.E. (2003). Eldanasaccharina (Lepidoptera: Pyralidae) in sugarcane(Saccharum hybrids), sedge (Cyperusdigitatus) and bulrush (Typha latifolia) in south-eastern Zimbabwe. Proc. S. Afr. Sug. Technol. Ass., 77: 266–274.

McFarlane, S. A., Govender, P. and Rutherford, R.S. (2009). Interactions between Fusarium species from sugarcane and the stalk borer Eldanasaccharina Walker (Lepidoptera: Pyralidae). Ann. Appl. Biol. Accepted 21 May.

Meyling, N. V. and Eilenberg, J. (2007). Ecology of the entomo pathogenic fungi Beauveriab assiana and Metarhiziumanisopliae in temperate agroeco systems: potential for conservation biological control. Biol. Control., 43: 145–155

Munir, M., Khan, W. A., Iqbal, J. and Ali, Z. (2014). Screening of advanced clones of NUVYT for resistance against sugarcane borers. Annual Program of Research Work., 2014: 55-57.

Ngi-Song, A. J., Overholt, W. A. and Stouthamer, R. (1998). Suitability of Busseolafusca and Sesamiacalamistis (Lepidoptera: Noctuidae) for the development of two populations of Cotesiasesamiae (Hymenoptera: Braconidae) in Kenya. Biol. Cont., 12: 208–214.

Rasmann, S., Kollner, T. G., Degenhardt, J., Hiltpold, I., Toepfer, S., Kuhlmann, U., Gershenzon, J. and Turlings, T. C. J. (2005). Recruitment of entomopathogenic nematodes by insect-damaged maize roots. Nature, 434: 732–737.

Schulthess, F., Caldwell, K.F. and Gounou, S.(2002). The effect of endophytic Fusarium verticillioides on infestation of two maize varieties by lepidopterous stem borers and coleopterangra in feeders. Phytopathol. 92: 120–128.

Sikandar, Z and Ahmad. N. (2021). Efficacy testing studies of different insecticides against borer pests in sugarcane crop. Annual Program of Research Work. 2021:46-48.

Smith, J. M., Conlong, D. E., Byrne, M. and Frerot, B. (2006). Response of Goniozusindicus (Hymenoptera: Bethylidae) to Sugarcane and Cyperus papyrus volatiles. Proc. S. Afr. Sug. Technol. Ass., 80: 250–255.

Smith, M. A., Wood, D. M., Janzen, D. H., Hallwachs, W. and Hebert, P. D. N. (2007). DNA barcodes affirm that 16 species of apparently generalist tropical parasitoid flies (Diptera, Tachinidae) arenotallgeneralists. Proc. Natl. Acad. Sci.USA, 104: 4967–4972.

Way, M. J., Leslie, G. W., Keeping, M. G. and Govender, A. (2006). Incidence of Fulmekiolaserrata (Thysanoptera: Thripidae) in South African sugarcane. Proc. S. Afr. Sug. Technol. Ass., 80: 199–201.

Zehnder, G., Gurr, G. M., Kuhne, S., Wade, M. R., Wratten, S. D. and Wyss, E. (2007). Arthropod management in organic crops. Ann. Rev. Entomol., 52: 57–80.

### INTEGRATED WEEDS CONTROL IN SUGARCANE RATOON MANAGEMENT WITH BIOTECHNOLOGICAL AND MOLECULAR APPROACHES

Salma Niaz\*, Abdul Khaliq\*, M. Shahzad Afzal\*, Naveed Akhtar\*\*, Waqas Raza Arshad\*\*\* \*Sugarcane Research Institute, Faisalabad \*\*Agronomic Research Institute (V&O), Faisalabad \*\*\*Sugarcane Research and Development Board, AARI, Faisalabad

#### ABSTRACT

Sugarcane crop is a cash and industrial crop contributing 0.7% in Pakistan's GDP. It is providing raw material for sugar mills operating in the country. The average cane yield in Punjab is 742mounds per acre. The progressive cane farmers is achieving more than 1500 mound per acre yield by growing latest varieties like CPF-249, HSF-240, CPF-234, CPF-250, CPF-251, CPF-252 and CPF-253 released by Sugarcane Research Institute, AARI, Faisalabad. Each variety has different features and needs different inputs and management requirements for plant and ration crop. Weeds management especially of narrow leaves is a difficult agronomic approach being faced in sugarcane. In most agriculture farmlands of sugarcane, weed management is predominantly reliant on herbicide application. Other agronomic methods and agro-technological manipulations were also being practiced for improving the productivity of sugarcane ratoons. It includes dismantling of ridges, stubble shaving, sub-soiling within rows, inter-culturing within rows and earthing up end of May. But these manipulations were adopted at small scale in farm area of sugar mills and few progressive farmers in Punjab. The weeds control is mainly done with use of weedicides of pre-emergence and post-emergence groups. However, the overuse and misuse of herbicides has resulted in the uptrend of herbicide-resistant weeds. Many biotechnological and molecular strategies can be focused on alterations of plant architecture, increased drought adaptation capabilities, increased salt tolerance, and increased pest and disease resistance and to reduce herbicide-resistant weeds. It is concluded that modern molecular approaches like Gene discovery, "omics," and genome editing technologies as a tool for current and future weeds management strategies in sugarcane plant and ratoon crop.

Keywords: Sugarcane, Weeds, Molecular, Biotechnological, Pakistan, Ratoon

#### INTRODUCTION

The world population is projected to increase from the current average of 7.6 billion people in 2020 to 8.6 billion people in 2030. The food security for increasing population great is а challenge for agriculture research meet the and demand of sugar of world population. One of the most significant challenges facing crop improvement programs globally is the capacity to adequately match crop

production with demand. thereby ensuring food security. Global crop production is affected by and various abiotic biotic stresses which are further worsened by climate change. Ratooning is ways of growing full cane crop from new growth of underground stubbles left in the field after reap of the plant crop (Singh et al., 2013). Ratoon crop is cost-effective for the farming communities of Pakistan because making cost is 30% less than plant crop with

saving of seed material as an extra benefit. It saves the cost of seedbed preparation, seed irrigation material. and planting labour due reduced crop period. In Punjab, half of sugarcane total area is engaged as ratoon (Naeem et al., 2019) but it contributes 30% to total cane production (Srivastava et al., 2012) due to improper attention of the farmers towards ratoons. Low vield of ratoon crop is primarily because of peculiarity ratooning potential of cultivated varieties (Rafig

2006) and pitiable et al.. ratoon management Techniques (Junejo et al., 2010). Good ratoon management practices and inherent ratoon potential of a variety is of prime importance for sustaining high cane and sugar productivity (Cheong and 2015). Teeluck. Vast acceptance of a variety depends very much on its ratooning potential (Verma, 2002). The sugarcane varieties will show good performance in ratoon crop only if accompanied with best techniques management (Hemwong et al., 2009). Otherwise, the variety will flop to perform in field (Singh and Singh, 2004). In world. sugarcane growing countries are taking two to five ratoons (Sundara et al., 2006). Good improvement of ratoon crop be determined by high sprouting of underground buds after harvesting of plant crop (Bashir et al., 2013). In multi-ratooning system. vield declined in successive ratoons can be enhanced by following good ratoon-management practices viz. loosening of inter-rows soil through chiseling, sub-soiling and earthing up to diminish soil compaction for root growth and preservation of trash to augment soil organic matter for resourceful utilization of water and nutrients (Hobbs et al., 2008). Furthermore, in ratoon sugarcane, the mortality of facultative tillers usually happens, especially in case those sprout from the aboveground uneven portions of canes left after harvest. Therefore, stubble shaving are recommended within a week of harvest of sugarcane (Ahmed and Giridharan 2000).

# Challenges in weed management

Despite the usefulness of integrated weeds

management (IWM), such strategies need to be heavily researched to determine the appropriate cultural, physical, and chemical methods that would be the most beneficial for the agro-ecological zone. Additionally, the change in the alobal climate has rendered some tried and true practices ineffective, leaving the door open to innovation in IWM. Climate change has raised complications in a number of different agricultural systems, and many of the challenges with weed management. Firstly, with the expected reduction in rainfall in already dry regions. the resilience of crops will be suffered. In this scenario. weeds have mechanisms to allow them to combat such stressors and out-compete the struggling crops, while also having extended periods of growth beyond their usual growing season (Ramesh et al., 2017). Weeds have ability quickly to accumulate mutations to be better adapted to rapidly changing climate scenarios, in contrast to many crops which rely on breeding programs to introgression desired traits in relatively slow manner. а Focusing more on the management side, climate change is expected to result in the need for new weed management strategies that need rapidly will to be implemented to be an effective combatant to the rapid climate variance. The change in climate will also result in the increased instability of current herbicides.

## Herbicide resistance in weeds

Continuous and non-judicious use of herbicides with the same mode of action creates herbicide resistance in weeds. From 1957 to 2020, the global reported number of unique cases of herbicideresistant weeds has increased from 2 to 507 2022). In general, (Heap. herbicide resistance mechanisms can be categorized into two broad target-site types: (1) resistance, and (2) non-target site resistance. Target-site resistance typically involves specific site mutations in the target enzyme, which prevents herbicide from binding to the target enzyme. Mutations could occur in the within binding sites the enzyme. Other forms of target-site resistance include target gene amplification (the increase in target gene copies) and the increase in target gene expression. These resistance mechanisms aim to increase the production capacity and abundance of the target in which higher enzyme, doses of a herbicide would be required to fully inhibit the target enzyme. Non-target site resistance stems from the physiological characteristics of the plant and how it absorbs, metabolizes, and/or sequesters the herbicide (Jugulam and Shyam, 2019). Another example of nontarget site resistance is through reducing translocation of the herbicide, so once the herbicide enters the source leaves they are prevented from reaching the and meristematic growing

tissues *via* the phloem and/or xylem. Reduced translocation can be due to sequestration, which traps the herbicide molecules within the source tissues, or altered activity of transporter proteins, which either prevent or limit the entrance of the herbicide molecules into the phloem and/or xylem.

## Weed seed bank persistence

Most weed species are known to be hardy and persistent in nature. producing thousands of seeds that can withstand various adverse environmental conditions. while staying dormant in the periods for long soil (Chauhan and Manalil, 2022). When optimal germination conditions are met, the seeds will germinate and compete with the crops sown on the same area of land. This makes weed management challenging. Seed dormancy is the main contributor to a persistent weed seed bank globally. It is a heritable genetic trait. Recent genetic and molecular studies on seed dormancy have provided important genomic information to aid the

understanding of seed dormancy in weeds.

#### Biotechnological and molecular approaches in weed management

Weeds are a detrimental global threat to crop production in both developing countries and developed (Chauhan, 2020). Overall, among the biotic factors causing crop losses, weeds contribute to the highest potential yield loss to crops. Some molecular approaches have been implemented in with herbicide conjunction application to reduce the proliferation of weeds in agricultural lands. Many molecular strategies for crop improvements have been largely focused on alterations of plant architecture, increased drought adaptation capabilities, increased salt tolerance, and increased pest and disease resistance. The development of glyphosateresistant crops enables the application of glyphosate, a non-selective herbicide, to eliminate unwanted weeds in the field at various application timings, thus enhancing the level of weed control (Masselet al., 2021).

Gene discovery, "omics," and genome editing technologies currently applied in crop research can be potentially applied to weeds as tools for weed management. Aside from GM methods, transient technologies relying on the non-transformative applications of **RNA** interference (RNAi) mechanism are also potential molecular approaches to control weeds instead of heavy reliance on herbicides.

These approaches could potentially manipulate expression of key genes in weeds to reduce its fitness and competitiveness, or, by altering the crop to improve its competitiveness or herbicide tolerance, by the molecular technologies in weed management. Genome editing may be used to improve crop resilience and adaptability to environments, various improve yields in suboptimal conditions. One such approach is the development of herbicide-resistant crops, the well-known such as Roundup Ready resistant crops (Barry et al., 1997).

#### CONCLUSION

It is concluded from above discussion that Biotechnological and Molecular techniques, like, genome editing, CRISPR/Cas9, gene drives, OMICS and RNAi technology, may be used for future molecular research on weed management as a tool for integrated weeds management in ratoon and plant sugarcane crop along with agronomical manipulation approaches. It will improve level of weeds control, higher cane and sugar yield.

#### REFERENCES

Ahmed, S.N., and S. Giridharan. 2000. Study on the influence of management practices on Sugarcaneratoon crop. Indian Sugar 49(10): 835–837.

Anonymous. 1970. Laboratory Manual for Queens Land Sugar Mills. (5th Ed.) Watson, Ferguson and Co. pp. 94-150.

Bashir S., N. Fiaz A. Ghaffar and F. Khalid. 2012. Ratooning ability of sugarcane genotypes at different harvesting dates. International Sugar Journal. 114(1360): 273-276.

Barry, G. F., Kishore, G. M., Padgette, S. R., and Stallings, W. C. (1997). Glyphosate-tolerant 5enolpyruvylshikimate-3-phosphate synthases. US Patent. 5633435.

Bashir S., M. Hassan, N. Fiaz, Z. Khan and Z. Ali. 2013. Ratooning Potential of different promising sugarcane genotypes at varying harvesting dates. Journal of Agricultural and Biological Science. 8 (05): 437-440.

Chauhan, B. S., and Manalil, S. (2022). Seedbank persistence of four summer grass weed species in the northeast cropping region of Australia. *PLoS One* 17:e0262288. doi: 10.1371/journal.pone.0262288.

Chauhan, B. S. (2020). Grand challenges in weed management. *Front. Agron.* 1:3. doi: 10.3389/fagro.2019.00003.

Cheong, L.R., and M. Teeluck. 2015. The practice of green cane trash blanketing in the Irrigated zone of Mauritius: effects on soil moisture and water use efficiency of sugarcane. Sugar Tech. doi:10.1007/s12355-015-0374-1.

Government of Pakistan (GOP). Pakistan Economic Survey, 2019-20. Ministry of Food, Agriculture and Livestock, Federal Bureau of Statistics, Islamabad.

Hobbs, P.R., K. Sayre, and R. Gupta. 2008. The role of conservation agriculture in sustainable Agriculture. Philosophical Transactions of the Royal Society B 363: 543–555.

Heap, I. (2022). The International Herbicide-Resistant Weed Database [Online]. Available at: https://www.weedscience.org/Pages/ChronologicalIncrease.aspx (Accessed Feb 3, 2022).

Hemwong, S., B. Toomsan, G. Cadisch, V. Limpinuntana, P. Vityakon, and A. Patanothai. 2009. Sugarcane residue management and grain legume crop effects on N dynamics, N losses and growth of sugarcane. Nutrient Cycling in Agroecosystems 83: 135–151.

Junejo SG, Kaloi M, Panhwar RN, Chohan M, Junejo AA, Soomro AF. 2010. Performance of Newly developed sugarcane genotypes for some qualitative and quantitative traits under ThattaConditions J. Plant Anim. Sci. 20(1):40-43.

Jugulam, M., and Shyam, C. (2019). Non-target-site resistance to herbicides: recent developments. *Plants* 8:417.

Massel, K., Lam, Y., Wong, A. C. S., Hickey, L. T., Borrell, A. K., and Godwin, I. D. (2021). Hotter, drier, CRISPR: the latest edit on climate change. *Theor. Appl. Genet.* 134, 1691–1709. doi: 10.1007/s00122-020-03764-0.

Naeem A, Khaliq A, Yasin M, Khursheed MR, Ahmad MF. 2019. Comparative evaluation of Ratooning potential of sugarcane clones. Int. J. Bio Res. 4(2): 07-09.

Ramesh, K., Matloob, A., Aslam, F., Florentine, S. K., and Chauhan, B. S. (2017). Weeds in a changing climate: vulnerabilities, consequences, and implications for future weed management. *Front. Plant Sci.* 8:95.

Rafiq, M., A. A. Chatta and M.R. Mian. 2006. Ratooning potential of different sugarcane genotypes under Faisalabad conditions. J. Agric. Res. 44(4): 269-275.

Singh, Dilip and S. M. Singh, 2004. Agro techniques for multiple ratooning in sugarcane. Indian Journal of Agronomy, 49 (4): 285-287.

Srivastava, T. K., M. Lal, and S. K. Shukla. 2012. Agronomic research in sugar crops: an overview. Indian Journal of Agronomy 57(3<sup>rd</sup> IAC Special Issue): 83–91.

Steel R. G. D., Torrie J. H. and Dickey D. A. 1997. Principles and procedures of statistics. A biometrical approach. 3rd Ed. McGraw Hill Book Co., Inc. New York, USA. pp. 400-428.

Sundara, B., 2006. Agro-technology for improving sugarcane productivity multi-ratooning. Cooperative Sugar, 37 (12): 37-52.

Singh, P., R. K. Rai, A. Suman, T. K. Srivastava, K. P. Singh, and R. L. Yadav. 2013. Ratooning Induced rhizosphere changes impede Nutrient acquisition and growth in sugarcane Ratoon crop during grand growth stage in sub-tropics. Sugar Tech 15(1): 52–64.

Verma, R. S. 2002. Sugarcane ratoon management, 202. Lucknow: International Book Distributing Co. Pvt. Ltd.

### REVIEW OF RECENT ADVANCEMENTS IN SUGARCANE GENOMICS

Muhammad Ehsan Khan Sugarcane Research and Development Board Corresponding Author email: <u>ehsansrdb@gmail.com</u>

#### ABSTRACT

Sugarcane is Pakistan's most important cash crop, primarily due to its industrial applications. In recent years, there has been a significant increase in sugarcane production, which has compensated for the decline in cotton production. Advances in sugarcane breeding have contributed to improving crop yield and agronomic traits, but further improvements are necessary to meet the rising demand for sugar and cope with changing climate conditions. However, conventional breeding methods have proven challenging due to sugarcane's complex and highly heterozygous genome. Despite these limitations, mapping techniques, genome-wide association studies, and genome editing techniques have identified key genes that enhance sugarcane's yield and resistance to disease and pests. Genome editing technologies, such as Zinc Finger Nucleotide (ZNF), Transcription activator-like effector nuclease (TALEN), and Cluster regularly interspaced short palindromic repeats (CRISPR/Cas associated protein 9), are efficient and precise tools that allow for rapid genome engineering. These technologies have been successfully applied in sugarcane to obtain new germplasm resources through gene-directed mutation. With whole genome sequencing data and knowledge of gene function, CRISPR-Cas 9 editing can rapidly generate new resources for key agronomic traits by precisely mutating important genes. This review focuses on the major differences and applications of these genome editing technologies in sugarcane plant engineering, emphasizing how they can facilitate molecular breeding and accelerate progress in basic research.

Keywords: Plant genome editing, crop improvements, Sugarcane breeding

#### INTRODUCTION

Sugarcane (Saccharum spp.) is a vital cash crop that is widely cultivated in countries with subtropical and humid climates such as the United States, India, Pakistan, China, Brazil, Australia, Cuba, and the Philippines. It is the primary source of global sugar production, accounting for 80% of the world's sugar supply. Sugarcane also yields various industrial high-end products like biofuel, waxes, and a diverse array of biofibers. There are two wild species (S. robustum and S. spontaneum) and four primarily cultivated species (S. edule, S. barberi, S. sinense, and S. officinarum) within the Saccharum genus. Modern sugarcane varieties genetically complex. are highly polyploid. and aneuploid, with 70%-80% of the genome composition from Saccharum officinarum and 10%-20% from S. spontaneum. Recently, Succharum spontaneum has emerged as a significant genetic resource for utilization in various sugarcane breeding programs, resulting in new polyploid and varieties with aneuploid chromosome counts ranging from 80 to 120. However. breeding conventional methods are challenging due to the narrow genetic pool and complicated genome of sugarcane.

Recent research on sugarcane has focused on molecular biology, including genome editing techniques, to achieve higher vields, increased sucrose content, and biotic and abiotic stress tolerance. These approaches benefit from the understanding of the complex interactions among genes, proteins, metabolites, and the genome but rely heavily on analytical methods such as bioinformatics and computational analysis. Significant progress has been made in understanding the molecular mechanisms of resistance sugarcane and tolerance to herbicides, cold, drought, and salinity stress, as well as plant development. Molecular marker approaches have elucidated the genome of modern structure sugarcane genotypes and derived phylogenetic relationships among the Saccharum complex. sugarcane Additionally. mapping genome helped experiments have detect marker-trait associations and validate the position of different essential genes.Top of Form

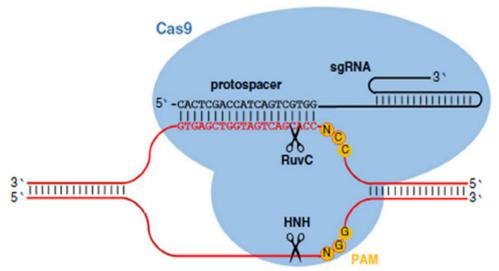
#### Gene-editing (GE) Technology: Gene-editing Tools:

Gene editing tools have revolutionized the field of biology by allowing for precise targeting of specific genes, which has opened up new opportunities for optimizing food production in

plants. The ability to generate permanent mutations or insertions using gene editing has made it possible to create knock-out phenotypes, which desirable in are most applications. Previous gene editing systems such as zinc finger nucleases and transcription activator-like effector nucleases were complex and time-consuming to use, but the Clustered Regularly Interspaced Palindromic Repeat (CRISPR)-associated protein system (CRISPR-Cas9) 9 offers a low-cost, simple, and efficient method for eukaryotic genome manipulation. CRISPR-Cas9 induces sitespecific double-strand breaks in genes of interest, which can be repaired through either non-homologous end joining or homology-directed repair pathway. Indels generated by NHEJ repair mechanisms disrupt the gene and create mutations, making

NHEJ more efficient than HDR for creating mutants.

The most widely used **CRISPR-Cas** system for editing genome is the CRISPR-Cas9 system, which consists of a single guide RNA and a Cas9 nuclease. The sgRNA binds to the target sequence and guides Cas9 to cleave the target The PAM sequence. sequence for the most **CRISPR**commonly used Cas9 system is 5'-NGG, and the sgRNA is usually driven by U6 promoters while the Cas9 gene expression is driven by CaMV or ubiquitin promoters. The CRISPRsystem Cas9 has been successfully used for genome editing in various plant species, and a platform called Plant Genome Editing Database has been created consolidate information to CRISPR/Casabout generated mutants of plant species.



**Figure-1:** The CRISPR/Cas9 system of Streptococcus pyogenes (Figure adapted from Puchta 2017, an Open Access article distributed under the terms and conditions of the Creative Commons by Attribution (CC-BY) license, <u>https://creativecommons.org/licenses/by/4.0/</u>])

#### Delivery of CRISPR Components:

CRISPR components can be introduced into plant different genomes via formats. including DNA, mRNA (in vitro transcripts or IVT), or protein. DNA format delivery allows for the use of various plant transformation techniques such as agroinfiltration. Agrobacterium infection. biolistics. electroporation, virus-based, **PEG-mediated** and transformation. The mRNA also efficient. format is leading to stable transgenic events with reduced off-target effects. Pre-assembled Cas9ribo-nucleoproteins gRNA (RNPs) can be directly delivered. eliminating the introduction of foreign DNA into the host genome. This has approach been successfully used in various including plants. tobacco, Arabidopsis, lettuce, rice. petunia. grapevine, apple, potato, maize, and wheat, as well as in rice zygote cells. Lipofection. а mammalian DNA transfer technique, has also been used to deliver preassembled Cas9-gRNA RNPs into tobacco plant protoplasts genome editing, for with Lipofectamine 3000 and RNAiMAX achieving optimal delivery efficiencies of 66% and 48%, respectively (Liu et al. 2020).

#### Editing Efficiency:

The efficiency of CRISPR-Cas9-mediated knock-out can be influenced by several factors, including the gene's location on the chromatin, with editing being more effective in euchromatin than in heterochromatin (Jensen et

al. 2017), the selection of sgRNA sites from the gene to be edited (Zhao et al. 2017), the number and length of sgRNAs used for a single gene knock-out (Zhang et al. 2016), the form of Cas9 used for delivery (i.e., DNA, mRNA, or protein) (Kouranova et al. 2016), and the expression threshold level of Cas9/sgRNA (Yuen al. et 2017). As а result. researchers typically select multiple sgRNAs for in vitro testing and then choose the one with the best editing performance and the least offtargeting effect for further in vivo work (Liang et al. 2016; Zhao et al. 2017).

#### Applications and Limitations of CRISPR Systems:

The CRISPR-Cas9 system has various applications knockouts, bevond gene including the elimination of an entire chromosome (Zuo et al. 2017) or gene at a specific locus(Srivastava et al. 2017), removal of unwanted plant selectable genes. replacement and repair of a dysfunctional allele(Li et al. 2018a, b), and generation of an opening for gene integration at a specific locus (Zhao et al. 2016; Begemann 2017). et al. However, CRISPR-Cas9-mediated sitedirected insertion is less efficient than CRISPR/Cas9mediated site-directed deletions. Additionally, CRISPR technology can target multiple loci simultaneously(Rozov et al. 2019), regulate gene expression through specific gene activation or suppression. and create

diversity genetic for breeding(Miao et al. 2018). The use of GE techniques can lead to the achievement of homozygotes with bi-allelic and mono-allelic mutations in the T0 generation, reducing breeding time significantly. Advanced imaging systems CRISPR/dCas9 based on have also been invented, such as fluorescence proteins fused to dCas9, which have been used to visualize telomere repeats in tobacco leaf cells. Plant breeders rely genetic diversitv on to improve elite cultivars, and the **CRISPR-Cas** system offers promising new opportunities to create genetic diversity in an unprecedented way. For example, researchers have used CRISPR-Cas to edit regulatory regions and elements. generate novel alleles with varying expression levels. and optimize inflorescence architecture in tomato. Additionally. advanced imaging systems based on CRISPR/dCas9 have been developed, and homozygotes with bi-allelic and mono-allelic mutations can be achieved in generation. the Т0 This reduces breeding time and accelerates the rate of crop improvement, particularly for polyploidy crops with multiple alleles. Furthermore, scientists have developed nucleic acid diagnostic kits for plant and human diseases using the unique properties of Cas12 and Cas13 nucleases, such as the SHERLOCKTM CRISPR kit, which is easy to use, fast, accurate, and fieldready. However, limitations of using the **CRISPR-Cas9**  system remain, including specific PAM sequence requirements that may limit target sites for Cas9 at certain loci and the large size of Cas9.

## Genome Editing and Sugarcane:

Jung and Altpeter (2016) reported the first successful genome editing of sugarcane using TALENs to knockout the COMT gene and reduce content. lianin Target mutations were achieved in 74% of transgenic lines, with up to 99% of wild-type COMT alleles mutated in various lines. However, transgenic events with 99% mutation frequency only resulted in a 29-32% reduction in lignin content. possibly due to biological plasticity. Nevertheless, Kannan et al. (2018) demonstrated that mutant lines in field conditions exhibited up to a 19.7% reduction in lignin content and saccharification increased efficiency without affecting biomass production. Recently, CRISPR-Cas9 technology used for has also been efficient and reproducible gene targeting in sugarcane (Eid et al. 2021; Oz et al. 2021), offering a simpler and more precise approach to editing complex polyploid However, genomes. high specificity and low off-target effects are still essential for effective genome engineering in sugarcane.

#### Prospects and Challenges of Genome Editing in Sugarcane:

Conventional breeding for developing sugarcane varieties is time-consuming and laborious. It can take up to 12-15 years for a breeding cvcle. and manipulating multiple genes or complex metabolic pathways is almost impossible. In contrast, the application of GE techniques, especially the CRISPR-Cas9 svstem. has manv advantages. This system can accurately generate precise mutations on both single and multiple genes in a shorter time. However, despite being various applied to plant CRISPR-Cas9species. mediated gene editing in sugarcane is limited due to of the lack genome sequencing and functional genomics studies on this crop. The recent availability of the early draft of sugarcane hybrid cultivar SP80-3280 and the monoploid reference genome of sugarcane, along with allele-defined genome sequencing, presents more opportunities for researchers **CRISPR-Cas** to utilize in systems sugarcane. However. the reference genome may not be enough as even a 1-bp difference between the real and the genome reference can prevent genome editing. Developing an efficient sugarcane transformation system for delivering GE components is also essential as sugarcane transformation efficiency is lower than other crops. Overexpressing maize developmental genes Bbm and Wus2 has shown to transformation increase efficiency in sorghum. sugarcane, indica rice callus, and non-transformable maize inbred lines. The use of a tissue-specific promoter ZmPLTP has been employed to generate fertile transgenic lines and avoid phenotypic abnormalities and sterility.

#### CONCLUSION

As the human population arows and available agricultural land remains limited. continuous crop improvement is necessary for food security. While conventional plant breeding techniques have been successful, they are slow and can take years to produce varieties. new Biotechnological methods, such as genome editing tools **CRISPR-Cas9** like the system, offer faster and more precise ways to improve crop vields. This technology genes for targets specific significantly modification, reducing the time required to produce varieties. new Although transgenic techniques have been successful in enhancing they remain crops, controversial and have led to negative public perception. As a result, the emergence of CRISPR-Cas9-based genome editing is becoming a more accepted option. New Breeding Techniques (NBT), CRISPR-Cas9, including considering them indistinguishable from traditionally bred plants. This is good news for biotech companies. breeders. and researchers using CRISPR to modify plants. Researchers have already used CRISPR-Cas9 to improve the quantity biofuel and quality of feedstocks such as algae and plant cell walls, as well as to engineer fermentation veasts

#### PSJ APRIL-JUNE, 2022 ISSUE

for more efficient bioethanol production. Sugarcane is a significant biofuel feedstock, and there has been substantial investment in quality improving its and quantity through transgenesis, limited with commercial We success. believe that genome editing will be used to identify important genes in sugarcane and their potential applications, particularly in bioenergy production. However, genetic improvement faces two

biological barriers: sexual reproduction in blooming plants and cellular equilibrium that ensures proper signaling between plant cells, tissues, ultimately and organs. affecting the plant's interaction with the environment. To overcome the former barrier, transgenic techniques can serve as a short-term solution, but it requires identifying the target modification gene for or introduction. Among the various breeding strategies, better gaining а

Vol. XXXVII, No.02

understanding the of biological network can effectively improve crop management, enhance the potential of existing crops, and facilitate the development of new ones through "assisted genomic selection." Utilizing biological systems genomics and tools can integrate knowledge from various fields and. with statistical analysis, lead to better management of new sugarcane varieties.

#### REFERENCES

Waclawovsky, A.J.; Sato, P.M.; Lembke, C.G.; Moore, P.H.; G.M. Souza(2010). Sugarcane for bioenergy production: An assessment of yield and regulation of sucrose content. Plant Biotechnol. J., 8, 263–276.

Ali, A.; Pan, Y.-B.; Wang, Q.-N.; Wang, J.-D.; Chen, J.-L.; S.-J.Gao(2019) Genetic diversity and population structure analysis of Saccharum and Erianthus genera using microsatellite (SSR) markers. Sci. Rep. 9, 395.

Zhang, J.; Zhang, X.; Tang, H.; Zhang, Q.; Hua, X.; Ma, X.; Zhu, F.; Jones, T.; Zhu, X.; Bowers, J. (2018). Allele-defined genome of the autopolyploid sugarcane Saccharum spontaneum L. Nat. Genet. 50, 1565.

Garsmeur, O.; Droc, G.; Antonise, R.; Grimwood, J.; Potier, B.; Aitken, K.; Jenkins, J.; Martin, G.; Charron, C.; C. Hervouet(2018). A mosaic monoploid reference sequence for the highly complex genome of sugarcane. Nat. Commun. 9, 2638.

Mustafa, G.; Joyia, F.A.; Anwar, S.; Parvaiz, A. M.S. Khan (2018). Biotechnological interventions for the improvement of sugarcane crop and sugar production. In Sugarcane-Technology and Research; IntechOpen: London, UK, pp. 113–138.

Barnabas, L.; Ramadass, A.; Amalraj, R.S.; Palaniyandi, M.; Rasappa, V. Sugarcane proteomics: An update on current status, challenges, and future prospects. Proteomics 2015, 15, 1658–1670.

Kaur, L.; Dharshini, S.; Ram, B.; Appunu, C. Sugarcane Genomics and Transcriptomics. In Sugarcane Biotechnology: Challenges and Prospects; Springer: Cham, Switzerland, 2017; pp. 13–32.

Sanghera, G.S.; Singh, R.P.; Tyagi, V.; Thind, K (2017). Recent genomic approaches for sugarcane improvement: Opportunities and challenges. In Quality and Quantum Improvement in Field Crops; Agrobios: Jodhpur, India, pp. 109–152.

Khueychai, S.; Jangpromma, N.; Daduang, S.; Jaisil, P.; Lomthaisong, K.; Dhiravisit, A.; Klaynongsruang, S. (2015). Comparative proteomic analysis of leaves, leaf sheaths, and roots of drought-contrasting sugarcane cultivars in response to drought stress. Acta Physiol. Plant. 37, 88.

Augustine, S.M.; Narayan, J.A. Syamaladevi, D.P. Appunu, C. Chakravarthi, M.; Ravichandran, V.; Tuteja, N.; Subramonian, N. (2015). Overexpression of EaDREB2 and pyramiding of EaDREB2 with the pea DNA helicase gene (PDH45) enhance drought and salinity tolerance in sugarcane (Saccharum spp. hybrid). Plant Cell Rep. 34, 247–263.

Li, M.; Liang, Z.; Zeng, Y.; Jing, Y.; Wu, K.; Liang, J.; He, S.; Wang, G.; Mo, Z.; Tan, F. (2016). De novo analysis of transcriptome reveals genes associated with leaf abscission in sugarcane

(Saccharum officinarum L.). BMC Genom. 17, 195.

Su, Y.; Xu, L.; Wang, Z.; Peng, Q.; Yang, Y.; Chen, Y.; Que, Y. (2016). Comparative proteomics reveals that central metabolism changes are associated with resistance against Sporisorium scitamineum in sugarcane. BMC Genom. 17, 800.

Allwood, J.W.; Ellis, D.I.; Goodacre, R. (2008). Metabolomic technologies and their application to the study of plants and plant–host interactions. Physiol. Plant. 132, 117–135.

Abudayyeh, O.O., J.S. Gootenberg, P. Essletzbichler, S. Han, J. Joung, J.J. Belanto, V. Verdine, D.B.T. Cox, M.J. Kellner, A. Regev, E.S. Lander, D.F. Voytas, A.Y. Ting, and F. Zhang(2017). RNA targeting with CRISPR-Cas13. Nature 550: 280–284.

Abudayyeh, O.O., J.S. Gootenberg, M.J. Kellner, and F. Zhang(2019). Nucleic acid detection of plant genes using CRISPR-Cas13. CRISPR Journal 2 (3): 165–171.

Amalraj, V.A., and N. Balasundaram (2006). On the taxonomy of the members of 'Saccharum complex.' Genetic Resources and Crop Evolution 53: 35–41.

Andersson, M., H. Turesson, N. Olsson, A.S. Fa<sup>"</sup>It, P. Ohlsson, M.N. Gonzalez, M. Samuelsson, and P. Hofvander(2018). Genome editing in potato via CRISPR-Cas9 ribonucleoprotein delivery. Physiologia Plantarum 164 (4): 378–384.

Araujo, K., D. Mahajan, R. Kerr, and M. da Silva(2017). Global biofuels at the crossroads: An overview of technical, policy, and investment complexities in the sustainability of biofuel development. Agriculture 7 (4): 32.

Augustine, S.M., J.A. Narayan, D.P. Syamaladevi, C. Appunu, M. Chakravarthi, V. Ravichandran, et al. 2015c. Overexpression of EaDREB2 and pyramiding of EaDREB2 with the pea DNA helicase gene (PDH45) enhance drought and salinity tolerance in sugarcane (Saccharum spp. hybrid). Plant Cell Reports 34: 247–263.

Balat, M., H. Balat, and C. O<sup>°</sup> z. 2008. Progress in bioethanol processing. Progress in Energy and Combustion Science 34: 551–573. Barrera, I., M.A. Amezcua-Allieri, L. Estupin<sup>°</sup>an, T. Martı nez, and J. Aburto. 2016. Technical and economical evaluation of bioethanol production from lignocellulosic residues in Mexico: Case of sugarcane and blue agave bagasses. Chemical Engineering Research and Design. 107: 91–101.

Baulcombe, D. 2004. RNA silencing in plants. Nature 431: 356–363. Begemann, M.B., B.N. Gray, E. January, G.C. Gordon, Y. He, H. Liu, X. Wu, T.P. Brutnell, T.C. Mockler, and M. Oufattole(2017). Precise insertion and guided editing of higher plant genomes using Cpf1 CRISPR nucleases. Scientific Reports 7 (1): 11606.

Belhaj, K., A. Chaparro-Garcia, S. Kamoun, and V. Nekrasov. 2013. Plant genome editing made easy: Targeted mutagenesis in model and crop plants using the CRISPR/Cas system. Plant Methods 9: 39.

Bensah, E.C., and M. Mensah(2013). Chemical pretreatment methods for the production of cellulosic ethanol: technologies and innovations. International Journal of Chemical Engineering (hindwi). 2013: 719607.

Bertrand, E., L.P.S. Vandenberghe, C.R. Soccol, J.C. Sigoillot, and C. Faulds. 2016. First generation bioethanol In. Green fuels technology: Springer International Publishing, Switzerland. Bewg, W.P., C. Poovaiah, W. Lan, J. Ralph, and H.D. Coleman(2016). RNAi downregulation of three key lignin genes in sugarcane improves glucose release without reduction in sugar production. Biotechnology for Biofuels 9: 270.

Bhowmik, P., E. Ellison, B. Polley, V. Bollina, M. Kulkarni, K. Ghanbarnia, H. Song, C. Gao, D.F. Voytas, and S. Kagale(2018). Targeted mutagenesis in wheat microspores using CRISPR/ Cas9. Scientific Reports 8: 6502.

Bilal, M., M. Saeed, I.A. Nasir, B. Tabassum, M. Zameer, A. Khan, M. Tariq, A.J. Mohamed, and T. Husnain(2015). Association mapping of cane weight and tillers per plant in sugarcane. Biotechnology and Biotechnological Equipment 29: 617–623.

Boerjan, W., J. Ralph, and M. Baucher(2003). Lignin biosynthesis. Annual Review of Plant Biology 54: 519–546. Boettcher, M., and M.T. McManus. 2015. Choosing the right tool for the job: RNAi, TALEN or CRISPR. Molecular Cell 58 (4): 575–585.

Bonawitz, N.D., and C. Chapple(2010). The genetics of lignin biosynthesis: Connecting genotype

to phenotype. Annual Review of Genetics 44: 337–363.

Bortesi, L., and R. Fischer(2015). The CRISPR/Cas9 system for plant genome editing and beyond. Biotechnology Advances 33: 41–52.

Botella, J.R. 2019. Now for the hard ones: Is there a limit on CRISPR genome editing in crops? Journal of Experimental Botany 70 (3): 734–737.

Burstein, D., L.B. Harrington, S.C. Strutt, A.J. Probst, K. Anantharaman, and B.C. Thomas. (2017). New CRISPR-Cas systems from uncultivated microbes. Nature 542: 237–241.

Callaway, E. (2018). CRISPR plants now subject to tough GM laws in European Union. Nature 560: 16.

Carrasco-Valenzuela, T., C. Mun<sup>o</sup>z-Espinoza, A. Riveros, R. Pedreschi, P. Aru's, R. Campos-Vargas, and C. Meneses(2019). Expression QTL (eQTLs) analyses reveal candidate genes associated with fruit flesh softening rate in Peach [Prunus persica (L.) Batsch]. Frontiers in Plant Science 10: 1581.

Carroll, D. (2011). Genome engineering with zinc-finger nucleases. Genetics 188: 773–782.

Carvalho-Netto, O.V., J.A. Bressiani, H.L. Soriano, C.S. Fiori, J.M. Santos, G. Barbosa, M.A. Xavier, G.A.L. Landell, and G.A.G. Pereira(2014). The potential of the energy cane as the main biomass crop for the cellulosic industry. Chemical and Biological Technologies in Agriculture 1: 20.

Cebrian-Serrano, A., and B. Davies(2017). CRISPR-Cas orthologues and variants: Optimizing the repertoire, specificity and delivery of genome engineering tools. Mammalian Genome 28: 247–261.

Chandel, A.K., S.S. da Silva, W. Carvalho, and O.V. Singh (2012). Sugarcane bagasse and leaves: Foreseeable biomass of biofuel and bio-products. Journal of Chemical Technology and Biotechnology 87: 11–20.

Char, S.N., A.K. Neelakandan, H. Nahampun, B. Frame, M. Main, M.H. Spalding, P.W. Becraft, B.C. Meyers, V. Walbot, K. Wang, and B. Yang. 2017. An Agrobacterium-delivered CRISPR/Cas9 system for high-frequency targeted mutagenesis in maize. Plant Biotechnology Journal 15 (2): 257–268.

Chen, F., and R.A. Dixon(2007). Lignin modification improves fermentable sugar yields for biofuel production. Nature Biotechnology 25: 759–761.

Cho, S.W., J. Lee, D. Carroll, J.S. Kim, and J. Lee(2013). Heritable gene knockout in Caenorhabditis elegans by direct injection of Cas9–sgRNA ribonucleoproteins. Genetics 195 (3): 1177–1180.

Cong, L., F.A. Ran, D. Cox, S. Lin, R. Barretto, N. Habib, P.D. Hsu, X. Wu, W. Jiang, L.A. Marraffini, and F. Zhang (2013). Multiplex genome engineering using CRISPR/Cas systems. Science 339: 819–823.

Cox, D.B.T., J.S. Gootenberg, O.O. Abudayyeh, B. Franklin, M.J. Kellner, J. Joung, and F. Zhang. 2017. RNA editing with CRISPR-Cas13. Science 358: 1019–1027.

Dreissig, S., S. Schiml, P. Schindele, O. Weiss, T. Rutten, V. Schubert, E. Gladilin, M.F. Mette, H. Puchta, and A. Houben. 2017. Live cell CRISPR-imaging in plants reveals dynamic telomere movements. Plant Journal 91: 565–573.

Eid, A., C. Mohan, S. Sanchez, D. Wang, and F. Altpeter(2021). Multiallelic, targeted mutagenesis of magnesium chelatase with CRISPR/Cas9 provides a rapidly scorable phenotype in highly polyploid sugarcane. Frontiers in Genome Editing 3: 654996. https://doi.org/10.3389/fgeed.2021.65499.

Endo, M., M. Mikami, and S. Toki(2016). Biallelic gene targeting in rice. Plant Physiology 170 (2): 667–677. FAOSTAT. 2016. Available at http://www.fao.org/faostat/ en/#data/QC

Fargioni, J., J. Hill, S. Plosky, and P. Hawthorne(2008). Land clearing and biofuel carbon debt. Science 319: 1236–1238.

Fu, Y., J.D. Sander, D. Reyon, V.M. Cascio, and J.K. Joung(2014). Improving CRISPR-Cas nuclease specificity using truncated guide RNAs. Nature Biotechnology 32: 279–284.

### ASSESSMENT OF SUGARCANE INTER CROPPING WITH DIFFERENT CROPS

Kanza Khan<sup>1</sup>, Muhammad Ehsan Khan<sup>2</sup>
1. University of Agriculture, Faisalabad.
2. Sugarcane Research and Development Board
Corresponding Author email: kanzakhan3346@gmail.com

#### ABSTRACT

The rapidly growing population of Pakistan needs to fulfill its food and nutrition requirements, and to achieve this, a collaborative strategy must be adopted to increase productivity by intensifying land use. Intercropping, which involves cultivating multiple crops in the same space simultaneously, is an advanced management practice that improves soil fertility and increases yield on a given piece of land by utilizing a mixture of crops with different abilities in rooting, canopy structures, height, and nutrient requirements. Intercropping is particularly beneficial for smallholder farmers in the sub-tropics, where intercropping sugarcane and legumes is widespread due to the legume's ability to address declining soil fertility. This review paper focuses on the role of intercropping systems in improving the growth, yield, and nutrient status of sugarcane in smallholder farms in semi-arid areas of Pakistan and other countries. The study discusses the different intercropping systems used in sugarcane and their effectiveness in increasing productivity, profitability, water use efficiency, and controlling weeds, pests, and diseases. The findings of this study will be useful for researchers involved in this field.

#### INTRODUCTION

Sugarcane, which belongs to the Saccharum spp. hybrid complex, is a significant cash crop in Pakistan and is widely grown tropical in and subtropical regions of the world. The sugar industry is second-largest the agroindustry in Pakistan, providing a source of food, fuel, fodder, and fiber, and plays a crucial role in the national economy. Globally, sugarcane is the main sugar-producing crop and contributes nearly 75% to the total sugar pool. In Pakistan, sugarcane covers an area of 1.260 million hectares, with a production of 88.65 million tonnes and a vield of 70.34 kg/ha. In Punjab, it covers 7.76 lakh hectares, with a production of 577 lakhtonnes and а productivity of 73.36 tonnes/ha (PSMA, 2021).

Intercropping was initially practiced as insurance against crop failure under rain-fed conditions. Nowadays, intercropping is mainly used to increase productivity per unit area and provide stability in production. The intercropping system efficiently utilizes resources and increases productivity. The primary advantage of intercropping is achieving greater yield on a given piece of land by making more efficient use of growth resources through the use of a mixture of crops with abilities, different rooting structures, canopy height, and nutrient requirements based on the complementary utilization of growth resources by the component crops. Legumes, when used as an increase intercrop. soil conservation through greater ground cover than sole cropping and improve soil fertility through biological nitrogen fixation compared to monoculture. Sugarcane has a slow growth rate at the initial stage with low leaf canopy, providing sufficient uncovered area for some crops to be grown. As a longduration and widely spaced crop, sugarcane offers good possibilities for growing earlymaturing intercrops to harness the potentiality of the environment and use natural resources increase to production and net profit per unit area per unit of time. These features offer а potential scope to intercrop relatively short-duration and quick-growing crops to exploit

land resources more efficiently.

#### Literature Review:

Literature reviews on various aspects of intercropping in sugarcane, including growth, yield, economics, quality, soil nutrient status, as well as physical. chemical. and biological properties of soil, were collected in January, 2021 through internet searches using Google's search engines worldwide. review encompassed The published and unpublished sources such as reports, research papers, and theses from the past 25 years.

**RESULT AND DISCUSSION** Yield Growth. and Economics of intercropping profitable The most intercropping combination for sugarcane is cane and garlic. Garlic does not compete much with sugarcane for light and shade, and a companion crop of these two plants resulted in a cane yield of 111.47 tha-1 and 4.18 tonnes of garlic, with only a slight decline of 5.3% in cane yield compared to the sole crop of sugarcane in Pakistan (Bukhtiar and Muhammad in 1988). Ali et al. in 1987 and Ahmed et al. in 1991 made similar observations. Garlic should be sown in between the rows of sugarcane at a spacing of 90 cm. То maintain proper spacing, three rows of garlic should be planted at a distance of 15 cm, with a plant-to-plant distance of 10 cm. If the cane row spaces are 120 cm wide, garlic can be planted in four rows (Ali et

al. in 1987 and Ahmed et al.

in 1991). According to Patel et al., (1984), intercropping sugarcane with garlic resulted in significantly higher yields of cane, single cane weight, and commercial cane sugar. Meanwhile. sugarcane intercropped with onion had higher intercrop yield and net return. In order to cultivate a profitable combination of cane and maize, it is essential to plant both crops early in the season, preferably during the first two weeks of February. This allows the maize to grow and mature rapidly before the tillering phase of the cane. For cane, inter-row spaces of 90 to 120 cm should be used for planting, and the inter-row spaces of the cane should be cultivated as a seed bed for maize when irrigated. The maize seeds should be drilled in a single row if the cane row space is 90 cm and in two rows if the space is 120 cm, with a plant-to-plant distance of 15 cm. If trench planting is used, maize seeds should be dibbled on both sides of the trenches. То avoid the exhaustive effect of this intercrop combination. the cane field should be enriched with a significant amount of farm yard manure. Dual row planting of cane can accommodate exhaustive crops and produce profitable yields of both cane and maize (Balde, 2011)Rana et al. (2006)observed that sugarcane + maize intercropping produced significantly higher millable cane and cane vield, with cane equivalent yield being the highest under this with treatment. along maximum net return and B:C ratio. However, sugarcane +

mash gave equally high yield. Varghese et al. (2006)revealed sugarcane that intercropped with vegetable peas produced significantly higher cane yield, land equivalent ratio (LER), and B:C ratio, with higher cane weight.Peas are a legume vegetable that provides a profitable return and can be used for good biomass incorporation into soil. Additionally, all leafy vegetables have high biomass that is beneficial for soil incorporation. To achieve successful intercropping of vegetables, wider rows are recommended, but paired row is preferable for planting managing profitable intercropping. It is important note that intercropping to should aim to supplement cash returns without compromising cane yield. Singh et al. (2010) found that single-bud vertical planted garlic sugarcane + had significantly higher cane yield and cane equivalent yield, followed by sugarcane + radish vegetable, with net return and B: C ratio being the higher in former treatment. Numerous studies have investigated the impact wheat of cane and intercropping systems on yield, with results crop showing that wheat has a negative effect on cane yield. For example, one study found that while a sole cane crop produced 133.97 tons per hectare, a combination of and wheat vielded cane 118.04 tons of cane and 3.385 tons of wheat per hectare, resulting in a 11.9% reduction in cane vield (Bukhtiar and Muhammad,

1988).Suryawanshi et al. (2010) reported that sugarcane + wheat intercropping gave higher net monetary return (NMR) and B: C ratio.

According to Islam and Islam (2016), the cane + potato combination is the most profitable intercropping system, provided that planting time, fertilizer needs, weed control. and earthing up operations carefully are managed. In particular, September-planted potato on ridges followed by cane in furrows produced higher yields than potato alone in September followed by cane in March. Furthermore, the highest cane vield was achieved with a September cane planting at 90 cm (101.16 t ha-1) with potato ha-1), (10.45 t followed closely by cane at 120 cm potato (Malik with and Kamoka, 1992). Kumar et al. (2011)noticed that sole sugarcane and sugarcane + intercropping potato had similar cane yield, while sugarcane onion intercropping produced higher cane equivalent yield and net returns. Studies by Navyar et al. (1987) and Ahmed et al. (1988) shown that have intercropping okra and sugarcane highly is а profitable combination, with an EMV of more than one. Although there was а reported reduction in cane yield of 6 to 17%, the monetary return from the okra crop compensated for this vield decline (Table-13). To minimize the shading effect of intercrop, cane was the planted in dual row strips with row spacing of 45-135-45 cm.

Paired row planting with a spacing of 30-150-30 cm was found to be more financially beneficial than the 45-135-45 cm row spacing. Therefore, wider inter row spaces are recommended to reduce light and shade competition, and two adjacent cane rows at 30/45 cm can make up the required plant population for Keshavaiah et cane. al. (2014)reported that sugarcane + french bean gave similar yields to pure sugarcane while crop, sugarcane + bhendi had significantly higher cane equivalent yield, with sugarcane vegetable + soybean having higher total income and B: C ratio. Khippal et al. (2016) showed that sugarcane + pea intercropping had similar cane yield to sole sugarcane crop, with net return being higher in the former treatment. Rana et al. (2006) found that sugarcane + mash resulted in significantly higher juice sucrose levels, which were comparable to those of sole sugarcane, sugarcane + mustard, and sugarcane + maize. They also observed that this treatment produced the highest CCS. Inter-cropping with lentil crops can yield a reasonably good profit margin without affecting the productivity of cane. Lentil has minimal competition for light and nutrients, and does not shade cane, and can even improve cane yield through symbiotic nitrogen fixation (Akhter et al., 2001). In fact, an EMV of 1.33 and 1.20 was obtained from intercropping cane with lentil (Table-4). Furthermore, it has been observed that lentil

intercropped in ratoon can improve cane yield compared to a sole crop (Singh et al., 2008).Singh et al. (2008) revealed that the highest amounts of available nitrogen and soil infiltration rates were obtained from the sugarcane + lentil intercropping system in both plant and ratoon sugarcane, with the lowest bulk density also observed in this treatment. Singh et al. found significantly (2011)higher commercial cane sugar levels in sole which sugarcane. were comparable to sugarcane + LP (1:3), with significantly higher purity percentages observed in sugarcane + LP (1:4), but comparable to all treatments other except sugarcane + LG (B). Brix readings were also higher in the same treatment.

Patel and Patel (2012)observed significantly higher values of available nitrogen and available phosphorus in after sugarcane the soil harvest with the application of a 100% recommended dose of phosphorus with green aram intercrop treatment. while available K2O and S were found to be nonsignificant. Keshavaiah et al. (2014) reported significantly higher reducing sugar and ash levels in the sugar cane + French bean intercropping system, while sucrose levels were not significantly affected by various treatments.

#### CONCLUSION

Growing a variety of crops such as sunnhemp, maize, radish, linseed, pea, cucumber, wheat, soybean, onion, amaranth, green gram, returns. and french bean alongside intercrop sugarcane can increase enhance sugarcane yield while also improvin providing better economic physical

returns. In addition, this intercropping system can enhance soil quality by improving nutrient status and physical and chemical properties of the soil, resulting in better quality crops.

#### REFERENCES

Akhter, A., M. Afzal, M. Najeeb Ullah and A. A. Chattha (2001). Benefits of inter cropping lentil in autumn planted sugarcane at farmer's field. Pak. Sugar Journal. 16(4): 111-114.

Akhtar, M., Ahmed, R., 1999. Impact of various weed control methods on the productivity and quality of sugarcane. 767 Pak. J. Biol. Sci., 2, 217–219.

Ali, A., M. Arshad and S. Ahmed (1986). Effect of inter cropping moong bean and sunflower in sugarcane. J. Aqric. Res., 24 (2): 102-107

Ali, I. and K. Hussain (1987). Dextran-a yard stick for cane staleness. Proc. Ann. Conv. Pak. Soc. Sug. Tech.: 432-442.

Alam, M. J., Rahman, M. M., Islam, A. K. M. R., Hossain, M. S., Razzak, M. A., Rahman, M. S., Roy, H. P., & Islam, S. (2014). Productivity and profitability of onion seed crop-Mungbean sequential intercropping with sugar cane. Bangladesh J. of Sugarcane, Vol. (35), page 60–72.

Anjum, S.A., M. Raza, N. Hussain, M. Nadeem and N. Ali (2015). Studies on productivity and performance of spring sugarcane sown in different planting configurations. Am. J. Plant Sci., 6(19): 2984.

Azam, M. and M. Khan (2010). Significance of the sugarcane crops with special reference to Khyber Pakhtunkhwa. Sarhad. J. Agric. 26(2): 289- 295.

Asad, M., A. Rasool, M. Zubair, S. Hussain and S. Afghan (2013). Efficiency of different post emergency chemical applications for summer weeds managements in sugarcane. Proceedings of Pakistan Society of Sugar Technologists. Sept. 9-10, 2013, Rawalpindi.

Aslam, M., G. Mohammed and K.B Malik (1994). Economic feasibility of inter-cropping moong and maize under different inter row strips of sugarcane. Proc. Ann. Conv. Pak. Soc. Sug. Tech. 29: 128-134.

Aslam, M., G. Mohammed, M.A. Javed and K.B. Malik. (1997). Agro- economic studies in intercropping moong and soybean in sugarcane with varying sowing dates J. Agri. Res., 35 (16): 373-378.

Balde, A.B., Scope, L.E., Affholder, F., Corbeels, M., DaSilva, F.A.M., Xavier, J.H.V., Wery, J. Agronomic performance of no-tillage relay intercropping with maize under smallholder conditions in Central Brazil. Field Crop Res. 2011, 124, 240–251.

Bukhtiar, B.A and G. Muhammad (1988). Effect of planting patterns and sowing dates on cane yield and quality under Bahawalpur condition. J. Agric. Res., 26 (3): 181-187.

Bukhtiar, B.A. (1988). Comparative response of four sugarcane varieties to inter cropped wheat. J. Agric. Res., 26 (1): 35-39.

Bukhtiar, B.A. and G Muhammad, (1988). Feasibility of companion cropping with autumn planted cane. Pak. J. Agric. Res., 9 (3): 294-299.

Bukhtiar, S.M., G.C. Paul, M.A. Rashid and A.B.M. Rahman (2001). Effect of press mud and organic nitrogen on soil fertility and yield of sugarcane grown in high Ganges River flood plain soils of Bangladesh. Indian Sugar, L1: 235-240.

Bukhtiar, S.M., S. Roksana and A.Z.M. Moslehuddin (2015). Soil fertility and productivity of sugarcane influenced by enriched press mud compost with chemical fertilizer. SAARC. Jour. Agri. 13(2): 183-197.

Khippal,A., Singh,S.M., Singh,J.S., and Kumar,R. 2016. Legume research –an international journal, **39**(3): 411-418.

Kumar, S., Singh, S.S. and Singh, A. 2011. *Progressive Horticilture* **43**(1): 153-154.

Kumar, Sanjay., Rana, N.S., Singh, R. and Singh, Adesh. 2006. Indian Journal of Agronomy,

**51**(4): 271-273.

Nayyar, M.M., K.B. Malik, M.N. Kamoka and M.A. Gill. (1987). Economic feasibility of intercropping some field and vegetable crops in spring and autumn crop of sugarcane. Proc. Ann. Conv. Pak. Soc. Sug.Tech., 23:107-122.

Nazir, M.S., Jabbar, A., Ahmad, I., Nawaz, S., Bhatti, I.H (2001). Production Potential and Economics of Intercropping in Autumn-Planted Sugarcane. Int. J. Agric. Biol., (4): 140–142.

Nazir, M.S., M. Ahmad, G. Ali and M. Siddique (1985). Feasibility of intercropping berseem in autumn sown sugarcane planted in different patterns. Pak. J. of Agri. Res., Vol. 6 (4). 259-266.

Naik, Balaji. R., Rao, Mukunda., Ramanjaneyulu, A.V. and Sekhar, 2010. *Progresses Agriculture*,**10**: 244-246.

Naik, R. B., Rao, C. M., Ramanjaneyulu, A. V. and Sekhar, D. 2008. Prog. Agric., 8(2): 240-242.

Paul M. White Jr., G. Williams, H. P. Viator, R. P. Viator and Charles L. Webber (2020). Legume Cover Crop Effects on Temperate Sugarcane Yields and Their Decomposition in Soil. Agronomy. Vol. (10) page 703.

Patel, C.L., Patel, D.D. and Patel, M.N. (2007). Critical period of crop weed competition in sugarcane (Var. Co Lk 8001). Indian Sug., 6 (12): 27-32.

Patel, K. R., Vashi, R. D. and Damame, H. S. 1984. GAU Res. J., 10(1): 13-17.

Patel, V.M. and Patel, C.L. 2012. Agricultural Science Digest, 32(2): 117-122.

Rana, N. S., Kumar, S., Saini, S. K. and Panwar, G. S. 2006. Indian J. Agron., 51(1): 31-33.

### SUGAR INDUSTRY ABSTRACTS

Quantifying the effects of longevity of nitrification inhibitors on nitrogen losses in simulated sugarcane production MP Vilas, K Verburg, JS **Biggs and PJ Thorburn** Proceedings of the International Society of Sugar Cane Technologists, volume 30, 1179-1185, 2019

Nitrification inhibitors (NIs) are one form of enhanced efficiency fertiliser (EEF). They have the potential to reduce nitrogen (N) losses from sugarcane production systems. Their effectiveness in reducing N losses depends on the longevity of the inhibitory effect, which is, in influenced by their turn. persistence in an intact form in soils. Studies examining NIs have shown that their persistence can range between a few days to more than one year. In this study, we explore NIs of different persistence to assess their ability to reduce N losses for a case study in northern Queensland (Australia) bv modelling means of а approach. We found that an increase in the persistence of NIs, is correlated with a reduction in Ν losses. However, we also found that this reduction in N losses does not always translate into vield benefits. We examine specific cases some to illustrate the situations that result in yield benefits in the presence of NIs.

#### Screening of sugarcane for high nitrogen-use efficiency at the seedling stage

Yang Liu, Liao Fen, Muhammad Anas, Li Qiang, Peng Lishun, Huang Dongliang and Li Yangrui Proceedings of the International Society of Sugar Cane Technologists, volume 30, 1696–1702, 2019

experiment This screened high nitrogen-use efficiency (NUE) genotypes under low nitrogen pressure selection svstem. The important indexes affecting NUE of sugarcane were analysed. and the results provide a theoretical basis for the selection of high NUE in breeding and cultivation of sugarcane. In this study, seedlings of 58 sugarcane genotypes were evaluated in a hydroponic experiment with low-N (0.2 mmol/L N) and normal-N (2 mmol/L N) treatment. Growth. drv biomass and N accumulation distribution and characteristics in various sections of the plant were evaluated using descriptive statistic, principal component analysis and cluster analysis. The results indicated that morphology, biomass. nitrogen efficiency traits showed high genotypic variation for N treatments. Under low-N treatment, the dry weight of 58 genotypes varied from 0.64to14.75 g/plant, nitrogen accumulation varied from 5.53 to 63.00 mg/plant and

NUE varied from 115.4 to 279.3 g/g. Four factors contributed to 92.85% of the variance, according to 26 such as parameters dry weight and N uptake with N deficiency. Another five factors were contributed according to 19 82.21%, parameters under both normal and low N treatment nitrogen transfer such as coefficient and genetic potential. The data revealed that dry weight (whole plant, leaf, root), N uptake (whole plant. leaf. shoot). NUE (whole plant. leaf), leaf relative NUE, shoot relative drv weight, shoot relative N uptake and shoot genetic potential were the key factors involved in sugarcane high NUE. Fifty-eight sugarcane genotypes were clustered into four groups: high NUE group, partial high NUE group, partial low NUE group and low group.

# Value of the conversion of sugarcane-biomass xylans to alkyl glycosides

Narendra Mohan, Vishnu Prabhakar Srivastava and Anushka Agarwal Proceedings of the International Society of Sugar Cane Technologists, volume 30, 1833–1841, 2019

Alkyl glycosides are a class of biodegradable non-ionic surfactants with wide applications in cosmetics. detergent. food and pharmaceuticals. Commercial production of these surfactants is carried out in

multiple steps through Fischer glycosylation reactions extreme under conditions. Issues of economic sustainability and environmental concerns have attracted the attention of the sugar industry Indian to explore the feasibility valuing by-products biomass and chain development in a biorefinery concept. The Indian Sugar Industry crushes about 225-250 Mt of sugarcane each year, generating nearly lignocellulosic 40-44% of biomass residue, i.e. 90-100 Mt sugarcane bagasse and trash. Xylans are abundant polysaccharides in sugarcane biomass and remain underutilized to great а Finding extent. new applications for xylan-based molecules and value-added sustainable chemicals is of interest and challenging in developing economically viable industries based on sugarcane biomass. Our research was undertaken for selective transformation of sugarcane biomass-based xylans into surfactants. A one-step production of alkyl glycoside from sugarcane biomass has been developed bv direct. selective alcoholysis in fatty alcohol media under acid catalysis at moderate temperature in a short time. The fatty alcohol serves as both solvent and reagent. The reactions conditions were broadly investigated and good conversion of xylan was achieved. The alkyl glycoside produced was characterized by NMR, FT-IR and mass spectrometry. Results indicate that the synthesized compound exhibits interesting

surfactant properties for applications as ingredients for detergents. This provides a new paradigm for the use of sugarcane biomass as a raw material for renewable chemical industries and will contribute likelv towards economic sustainability of the sugar industry worldwide.

Optimization of a phenol typing system in sugarcane evaluate different to strategies against Diatraea saccharalis (Fabricius) F Budequer, MF Perera, G J Racedo. Michavila. G Gastaminza, MI Cuenya and AP Castagnaro Proceedings of the International Society of Sugar Cane Technologists, volume 30, 193–199, 2019

Diatraea Sugarcane borer saccharalis (Fabricius) (Lepidoptera: Crambidae) is the most important sugarcane pest in Tucumán, Argentina. Older larvae (L3, L4) bore into the stalks, disrupting the physiological integrity of the plant, facilitating fungi and bacteria colonization which indirectly reduce yield and quality of sugar. The aim of the present work was to optimize a sugarcane plantinfestation method with D. saccharalis under controlled conditions to help to evaluate different strategies to manage the pest. Different numbers of neonate larvae were placed in the leaf whorl of sugarcane seedling (2-months-old) of cultivars TUC 95-10, TUC 03-12 and LCP 85-384. 10 larvae of several instars were added in the liqule of second and third fully expanded leaves of single plants, 6-months old, of Vol. XXXVII, No.02 95-10. Trials were

TUC conducted under controlled conditions (28-30°C; 50-70%) Each RH). assav was repeated twice with 5-10 replicates treatment. per Several parameters were evaluated on 6-months-old plants: damaged sheath number. number of perforations in the internodes. total tunnel length, and deadheart symptom; whereas on sugarcane seedlings only the parameter last was measured. Dead-heart symptoms were observed on sugarcane seedlings in all treatments with neonate larvae. Sugarcane plants (6months-old) showed damage in the sheath and stem when infested with L2, L3 and L4 instars. whereas neonate larvae only produced sheath damage since they were unable to pierce and bore the sugarcane stem. Results suggested that different methods could be used to screen different strategies for sugarcane borer management.

#### Key considerations for high-performance continuous vacuum pans

BStC Moor, S Rosettenstein and N du Plessis Proceedings of the International Society of Sugar Cane Technologists, volume 30, 25–35, 2019

important The two most objectives for highа performance continuous vacuum pan (CVP) are good and crystal quality high exhaustions. То achieve these, the pan design needs to incorporate features that promote plug flow (a narrow crystal residence-time distribution), a high heattransfer coefficient (HTC) and vigorous circulation. Focussing on these will also achieve an energy efficient pan that can operate on a low steam-massecuite

temperature differential. Good plug flow is an essential for good crystal quality (low CV), which enables good purging with minimal washing and good exhaustions. This in turn minimises reboiling and its associated energy and sucrose losses. Low CVs also aid affination and for this frequently reason are raw-sugar included in specifications. Measures to achieve good plug flow include a good circulation profile and good intercompartment massecuitetransfer arrangements. Achieving plug flow has the greatest presented challenge to CVP designers, but the problem is shown to have been mastered in some horizontally-configured vertical tube pans. Key requirements high for exhaustions are appropriate seed supply, good feed (supersaturation) control. vigorous circulation and a high final Brix. Various ways in which circulation can be promoted are described. Recent information is that, contrary to previous belief, longer tubes perform as well as or better than shorter tubes in CVPs. Types of pans which do or do not have the desired attributes are mentioned. It is concluded that the type of pans best suited to meeting most of these requirements are

vertical tube CVPs, at least one type of which is shown to come close to achieving true continuous spiral massecuite flow.

Design and implementation of a web-based knowledge management system in the Sugarcane and Byproducts **Development** Company in Iran Babak Radmehr and Elena Baninemeh Proceedings of the International Society of Sugar Cane Technologists, volume 30, 831-839, 2019

The Sugarcane and Bvproducts Development Company includes seven agri-industry companies, with similar organizational structure and 18,000 skilled and experienced employees that assist in producing half of the sugar that is consumed in Iran. Because of its size and spread, it is necessary to have knowledge а management (KM) system to broadcast and share personal experiences and proficiency order improve in to performance. organizational The advantages of a KM system are in reducing redundancies, avoiding duplication of mistakes. preserving employee knowledge before leaving the promoting organization, successful experiences at all levels. and documentina explicit and implicit knowledge of personnel anywhere and anytime. The KM system in this company is based on the integration of two models - the Milton and Heisig models. A specific web application was designed and

implemented based on the proposed model that was integrated with a set of technologies, server-side and client-side programming languages. Frameworks and databases support the four main KM stages (discovery, capture. sharing and application). In the process of discovery, an efficient scoring system was implemented to motivate all personnel to participate, the performance of which was based on the algorithm defined according to importance of the user's activities. During the process of data capturing and sharing, it was established that the existing content management systems (CMS) did not meet the requirements of the company. resulting in implementation of a specific CMS from the outset. The CMS is an interface that allowed users to publish content directly to the Webbased KM system. This paper describes the implementation of a KM system in the Sugarcane Development Company.

Development of a further transgenic sugarcane cultivar resistant to glyphosate herbicide J Racedo, F Budeguer, MF Perera, MJ Soria Femenías, SN Ovejero, MI Cuenya, AP Castagnaro and AS Noguera Proceedings of the International Society of Sugar Cane Technologists, volume 30, 515-521, 2019

A previous development of transgenic glyphosateresistant sugarcane suitable for commercial release was carried out through genetic

horizontally-configured

transformation of cultivar RA 87-3. During the time elapsed to develop this technology, cultivars new commercial the produced by local breeding program have been adopted by farmers. The complex genetics of modern sugarcane cultivars, which interspecific are hybrids, highly polyploid and frequently aneuploid, make introgression the bv backcrossing of the transgene into other varieties extremely difficult. Direct transformation of new commercial cultivars or promising clones at final stages of a breeding scheme could greatly improve the development and adoption of transgenic sugarcane by both farmers and millers. The aim of this study was to obtain glyphosate-resistant transgenic events of the two recently released cultivars, TUC 95-10 and TUC 03-12, through the introduction via

microprojectile bombardment of plasmids harbouring the epsps and nptll genes. A total of 23 and 8 independent bombardments experiments were carried out on TUC 95-TUC 10 and 03-12. The respectively. stable transformation and integration of both epsps and nptll genes were determined by using PCR with specific primers. Transgenic events were evaluated for glyphosate tolerance under ex vitro conditions and for genetic similarity with donor plant by using target region amplification polymorphism (TRAP) molecular markers. Although plantlets of both varieties regenerated from calli, TUC 95-10 showed a

low tissue culture-response since only two events were obtained, whilst a total of 22 plantlets regenerated for TUC 03-12. One line of TUC 95-10 and three lines of TUC 03-12 were PCR-positive for both genes and different levels of herbicide-tolerance were observed. Genetic analyses of 213 TRAP loci from transgenic lines derived from TUC 03-12 indicate that two 99.30% events show of similarity with the nontransformed TUC 03-12 control and the third had 98.7% similarity. These are results encouraging as, based our on previous experience, the molecularmarker data suggest that the events are practically identical to their parental cultivar, and are suitable for future comparative field testing.

Multilocus Sequence Analysis highlights genetic diversity of Acidovorax avenae strains associated with sugarcane red stripe PD Fontana, N Tomasini, CA Fontana, V Di Pauli, PS Cocconcelli, GM Vignolo and SM Salazar Proceedings of the International Society of Sugar Cane Technologists, volume 30, 1728–1735, 2019 Pathogenic species of Acidovorax cause economically important diseases in monocotyledonous and dicotyledonous crops, including sugarcane, corn. millet. rice, oats, foxtail, watermelon and orchids. Sugarcane red stripe, caused by Acidovorax avenae, is present in the main

production areas around the world. In Argentina, red stripe affects about 30% of stalks milled with important economic losses when infections severe occur. MLST was used to explore the genetic diversity of this bacterium associated with red stripe in Argentina, as well as their phylogenetic The relationships. MLST analysis included sequences of from total 118 а Acidovorax, 15 A. avenae strains isolated from Argentina sugarcane production areas, A. citrulli from (93) melon and watermelon, A. avenae (9) from rice, millet, corn, vasey grass and sorghum, and A. oryzae (1) from rice. MLST analysis revealed five novel sequence types (STs) for the sugarcane A. avenae strains, constituting a clonal complex with a common and close When origin. genetic relationships with other Acidovorax were explored, strains sugarcane were related to A. avenae from other hosts and more distantly to A. citrulli. Signals of frequent recombination in several lineages of A. avenae detected were and we observed that A. oryzae is closely related to A. avenae strains. This study provides valuable data in the field of epiphytological and evolutionary investigations of A. avenae strains causing sugarcane red stripe. Knowledge of the genetic diversitv and host-strain specificity are important to select the genotypes with the best response to red stripe disease.

### **INTERNATIONAL EVENTS CALENDAR**

#### **2022 CONFERENCES & MEETINGS**

February 16-19, 2022	7th IAPSIT International Sugar Conference & Sugarcon-2022 "Sustainability of the Sugar and Integrated Industries: Issues & Initiatives" Indian Institute of Sugarcane Research
April 17-19, 2022	S.I.T. Orlando Conference Sugar Industry Technologists Orlando, Florida, USA
April 19-22, 2022	Australian Society of Sugar Cane Technologists Conference Australian Society of Sugar Cane Technologists Mackay MECC, Queensland, Australia
June 14-16, 2022	ASSCT Annual Florida & Louisiana Joint meeting American Society of Sugar Cane Technologists Hyatt Regency Coconut Point Bonita Springs FL USA
June 20-24, 2022	XVI International Congress on Sugar and Cane Derivatives: Diversification 2022, AZCUBA Sugar Group, the Cuban Association of Sugar Technicians and the Cuban Institute of Research on Sugarcane Derivatives Cuba
June 30-July 1, 2022	Conferencia Bonsucro México 2022 Bonsucro Mexico City, Mexico
July 29- August 3, 2022	American Sugar Alliance Symposium American Sugar Alliance Seattle, WA, USA Location TBC
August 16-18, 2022	94th SASTA Congress 2022 SASTA – South African Sugar Technologists' Association ICC, Durban, 45 Bram Fischer Rd, Durban, 4001 South Africa
August 16-19, 2022	28ª Feira Internacional da Bioenergia Fenasucro & Agrocana Centro de Eventos Zanini, Sertãozinho, Brazil
September 12-16, 2022	XII Congreso Tecnicaña (XII Tecnicaña Congress) Tecnicaña
November 22-23, 2022	31st ISO International Seminar, International Sugar Organization

### **GUIDELINES FOR AUTHORS**

#### Dear Fellow Author(s),

Pakistan Sugar Journal (PSJ) offers research, analysis, and reviews to keep its local and international readership up to date with latest developments in the sugar industry. PSJ takes into account the application of research and focuses on areas in agriculture related to sugar, milling and processing.

In order to have your articles published in the PSJ, you are requested to adhere to the below instructions and prerequisites to enable timely review of your submissions by the editorial board:

- I. Write the title of your article in CAPITAL LETTERS in the center of the page.
- II. Write the complete name of all authors with their addresses it is compulsory in the text. References should be cited by author and years as, for one, two or more authors (Hammer, 1994, Hammer and Rouf, 1995; Hammer *et al.*, 1993), respectively.
- III. Write HEADINGS in bold letters and in the center of the page.
- IV. Type your article only in ARIAL format.
- V. Send TABLES and FIGURES on separate page with bold title and mark its numbers correctly.
- VI. Observe the following rule for REFERENCE, for one author: Hussain, K. 1991 for two authors; Khan, M. and A. Habib 1995, for more than two; Ali, K., A. Hussain and S. Nasir, 1990.
- VII. Always send two soft copies and one hard copy of CD. Please do not use FLOPPY DISK for this purpose.
- VIII. Send copies on an A-4 size page, preferable LASER PRINT in word document
- IX. Papers published in the PSJ are free of charges (for authors).
- X. Send your papers to following address by mail or email:

#### Dr. Shahid Afghan

Editor-in-Chief, Pakistan Sugar Journal Shakarganj Sugar Research Institute, Jhang (Pakistan) Phone: +92 47 111-111-765 | Ext. 602, 603 Email: shahid.afghan@shakarganj.com.pk.